

## Review

# Primary atopic disorders: inborn errors of immunity causing severe allergic disease

Maryam Vaseghi-Shanjani<sup>1,2</sup>, Simran Samra<sup>1,2</sup>, Pariya Yousefi<sup>1</sup>, Catherine M Biggs<sup>1</sup> and Stuart E Turvey<sup>1,2</sup>



Allergic diseases, including asthma, allergic rhinitis, atopic dermatitis, and food allergies, are driven by dysregulated immune responses, often involving IgE-mediated mast cell and basophil activation, Th2 inflammation, and epithelial dysfunction. While environmental factors are well-known contributors, the genetic components underpinning these conditions are increasingly understood. Traditionally viewed as polygenic multifactorial disorders, allergic diseases can also be caused by single-gene defects affecting the immune system and skin epithelial barrier, leading to profoundly dysregulated allergic responses. These monogenic allergic disorders are collectively referred to as primary atopic disorders or PADs. To date, over 48 single-gene defects have been established to cause PADs. This review highlights (i) the significance of PADs, (ii) the biological pathways involved in the pathogenesis of PADs, (iii) clinical strategies to differentiate PADs from their much more common polygenic counterparts, and (iv) diagnostic strategies for PADs.

## Addresses

<sup>1</sup> Department of Pediatrics, British Columbia Children's Hospital, The University of British Columbia, Vancouver, BC, Canada

<sup>2</sup> Experimental Medicine Program, Department of Medicine, The University of British Columbia, Vancouver, BC, Canada

Corresponding author: Turvey, Stuart E ([sturvey@bccchr.ca](mailto:sturvey@bccchr.ca))

**Current Opinion in Immunology** 2025, **94**:102538

This review comes from a themed issue on **Allergy and Hypersensitivity**

Edited by **Rod Rahmi** and **Carolyn Sokol**

For complete overview of the section, please refer to the article collection, "[Allergy & Hypersensitivity \(October 2024\)](#)"

Available online 27 February 2025

<https://doi.org/10.1016/j.coi.2025.102538>

0952-7915/© 2025 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## Single-gene defects as the cause of allergic disease

While the vast majority of people with allergies have a polygenic multifactorial disorder caused by the interaction of multiple genes with environmental exposures, we

have come to appreciate that allergic diseases can also be caused by single-gene defects. These monogenic disorders that affect the development, function, and regulation of the immune system and the skin epithelial barrier lead to dysregulated pathogenic allergic effector responses [1–3].

Collectively referred to as ‘monogenic allergic diseases’ or ‘primary atopic disorders’ (PADs), these Mendelian disorders are clinically and genetically heterogeneous with over 48 known genetic causes identified to date (Table 1) [4–6]. PADs represent a subset of the larger group of monogenic immune disorders collectively known as inborn errors of immunity or IEIs. Some PADs result from damaging variants<sup>1</sup> in genes that encode components of the immune system and the skin epithelial barrier, such as cytokines, receptors, structural proteins, and enzymes. Other PADs are caused by variants that disrupt genes involved in immune system development, activation, or differentiation, such as transcription factors and signaling molecules (see Table 1) [5].

The clinical and molecular heterogeneity of PADs underscores their complexity and emphasizes the need to understand the mechanisms involved in their pathogenesis. This understanding is crucial to develop targeted therapeutics that can effectively address the underlying molecular causes of these conditions. This review aims to highlight the significance of PADs, the disrupted biological pathways involved in their pathogenesis, and the central role clinical genetic sequencing plays in advancing affected patients to diagnosis and treatment.

## Categorizing the disrupted cellular and molecular pathways that cause primary atopic disorders

PADs are driven by genetic variants that interfere with the normal development and function of the immune system, the integrity of epithelial barriers, or cellular signaling [1]. Variants in these genes impact an array of cellular mechanisms that lead to the disruption of normal

<sup>1</sup> Throughout this Review, we will use the term ‘damaging variant’ to describe a disease-causing genetic change, rather than ‘mutation’ which has potentially pejorative connotations.

Table 1

Pathogenic mechanisms and an illustrative list of genes implicated in monogenic allergic diseases.

Pathogenic mechanism	Genes
Disruption of skin barrier function	<i>FLG</i> , <i>SPINK5</i> , <i>CDSN</i> , <i>DSG1</i> , <i>DSP</i> , <i>CARD14</i>
Granulocyte dysregulation	<i>KIT</i> , <i>PLCG2</i> , <i>ADGRE2</i>
Abnormal actin cytoskeleton remodeling	<i>WAS</i> , <i>WIPF1</i> , <i>ARPC1B</i> , <i>DOCK8</i> , <i>NCKAP1L</i> , <i>CARMIL2</i> , <i>STK4</i> , <i>MSN</i>
Attenuated antigen receptor signaling	<i>CARD11</i> , <i>CARD14</i> , <i>MALT1</i> , <i>CARMIL2</i> , <i>RFXANK</i>
Abnormal T cell development and/or Restriction of the T cell receptor repertoire	<i>LIG4</i> , <i>DCLRE1C</i> , <i>RAG1</i> , <i>RAG2</i> , <i>ADA</i> , <i>IL7RA</i> , <i>IL2RG</i> , <i>ZAP70</i> , <i>CHD7</i> , <i>TBX1</i> , 22q11.2 deletion syndrome, <i>FOXP3</i> , <i>TBX21</i> , <i>IKZF1</i> <sup>GOF</sup>
Altered cytokine signaling	<i>JAK1</i> <sup>GOF</sup> , <i>STAT1</i> <sup>GOF</sup> , <i>STAT3</i> <sup>DN</sup> , <i>STAT3</i> <sup>GOF</sup> , <i>STAT5B</i> <sup>LOF</sup> , <i>STAT5B</i> <sup>GOF</sup> , <i>STAT6</i> <sup>GOF</sup> , <i>IL2RA</i> <sup>LOF</sup> , <i>IL4RA</i> <sup>GOF</sup> , <i>TGFBR1</i> , <i>TGFBR2</i> , <i>ERBB2IP</i> , <i>IL6ST</i> , <i>IL6R</i> , <i>ZNF341</i> , <i>FOXP3</i> , <i>TBX21</i>
Altered cellular metabolism	<i>PGM3</i> , <i>CARD11</i> , <i>MALT1</i>

NB: due to their pleiotropic functions, some genes are listed under multiple pathogenic mechanisms.

immune cell functions and/or skin integrity, ultimately resulting in unchecked allergic inflammation [1,3]. These genetic variants can cause disease through the full range of genetic mechanisms, including gain-of-function (GOF), loss-of-function (LOF), and dominant-negative (DN) effects, each contributing to disease pathology through distinct modes of inheritance, including autosomal dominant, autosomal recessive, X-linked, and *de novo* disease. In the following paragraphs, we will group the individual PADs into mechanistic categories, and a more comprehensive list of currently well-characterized PADs is presented in Table 1. Recognizing the pleiotropic nature of many genes, some PAD genes are listed under more than one category: for example, biallelic disruption of *TBX21* causes deficiency of the transcription factor T-bet, affecting both the development of T cells (and many other immune cells) while also causing allergic inflammation due to excessive T helper 2 (Th2) cytokine production by adaptive CD4<sup>+</sup> αβ T lymphocytes [7–9].

### Disruption of skin barrier function

LOF variants in proteins that maintain the integrity of the epithelial barrier are a well-established cause of PADs. The integrity of the epithelial barrier is maintained by structural proteins like flaggrin (encoded by *FLG*), protease inhibitors such as serine peptidase inhibitor Kazal type 5 (*SPINK5*), and proteins involved in intercellular adhesion including corneodesmosin (*CDSN*), desmoglein 1 (*DSG1*), and desmoplakin (*DSP*). Germline LOF variants in these proteins impair skin barrier function resulting in enhanced exposure of antigen-presenting cells to environmental antigens, penetration of microbiota, and water loss [1,10]. These lead to the secretion of alarmins, such as interleukin (IL)-25, IL-33, and thymic stromal lymphopoietin, which are key regulators of type 2 immunity and trigger an allergic response [11]. Allergic inflammation persists because of downstream type 2 cytokines, such as IL-4 and IL-13 [12]. Although here we discuss the genes that are implicated in skin barrier function, it is also important to note that barrier disruption in the pathogenesis of atopy

may not be limited to skin — the disruption of the gastrointestinal tract or even respiratory mucosa could conceivably contribute [13]. For instance, in a recent study, Laky et al. demonstrated that intrinsic defects in transforming growth factor beta (TGF-β) receptor signaling within the gut epithelium can independently drive eosinophilic esophagitis, highlighting the role of epithelial cells not only as a physical barrier but also as key regulators of local immune responses through altered differentiation and alarmin production [14].

### Granulocyte dysregulation

Dysregulation of granulocytes, encompassing basophils, eosinophils, mast cells, and neutrophils, is another mechanism leading to heightened type 2 inflammation and development of PADs [15]. In healthy individuals, granulocytes (classically mast cells) release proinflammatory mediators after sensing an allergen through their high-affinity immunoglobulin E (IgE) receptors. However, dysregulation of granulocytes through a variety of GOF mechanisms results in PADs. For example, variants in *KIT* result in mastocytosis due to uncontrolled mast cell proliferation. Beyond proliferation defects, abnormal mast cell degranulation can cause urticaria in these conditions [16]. Additionally, *PLCG2* variants implicated in cold-induced urticaria lead to constitutive activation of PLCγ2 at lower temperatures, triggering mast cell degranulation and histamine release [17]. Also, vibratory urticaria caused by variants in *ADGRE2* results from mechanical stimuli disrupting receptor subunit interactions, leading to mast cell activation and urticaria (or hives) [18].

### Abnormal actin cytoskeleton remodeling

As key components of the adaptive immune system, lymphocytes also play a pivotal role in the development and progression of allergic inflammation in PADs. The actin cytoskeleton is critical to lymphocyte function, playing a key role in T cell receptor (TCR) signaling, clonal expansion, and regulatory T cell (Treg) function. The actin cytoskeleton facilitates the movement and organization of signaling molecules at the immunological

synapse, allowing effective TCR signaling and activation. Additionally, actin dynamics are crucial for the proliferation and differentiation of T cells during clonal expansion, which is necessary for mounting an effective immune response. Tregs, which help maintain immune tolerance and prevent excessive immune responses, also depend on the actin cytoskeleton for their suppressive functions. LOF variants in proteins that regulate actin polymerization, such as those encoded by *WAS*, *WIPF1*, *ARPC1B*, *DOCK8*, *NCKAP1L*, *CARMIL2*, *STK4*, and *MSN* characteristically lead to severe allergic inflammation, recurrent infections, and loss of immune tolerance to allergens and self-antigens [19–24]. The exact mechanisms by which each of these variants drives allergic inflammation are not entirely understood; nonetheless, several key insights have emerged. For instance, *DOCK8* deficiency skews CD4<sup>+</sup> cells towards a Th2 and away from a Th1 bias, indicating a T cell–intrinsic mechanism [25]. Additionally, these variants are linked to Treg cell dysfunction, further contributing to skewed Th2 responses that go unchecked and the concomitant autoimmunity observed in these disorders [26].

#### Attenuated antigen receptor signaling

Attenuated signaling through the TCR has also been linked to the development of Th2 inflammation in PADs [9]. Low-avidity interactions between a TCR and an antigen-derived peptide presented on a major histocompatibility complex (MHC) molecule promote naïve T cells to differentiate into Th2 cells [27]. Low-avidity interactions can be caused by variants in proteins that regulate the expression of MHC class II molecules (*RFXANK*) [28] or facilitate TCR signal transduction (*CARD11*, *MALT1*, and *ZAP70*; see Table 1 for a complete list). *CARD11* and *MALT1* are components of the CBM signalosome complex, and together they regulate the activation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) signaling pathway following TCR activation, which results in the induction of proinflammatory genes. Impairment in NF-κB signaling skews the cytokine milieu towards a Th2 phenotype, upregulating Th2 signaling and inflammation through several mechanisms [29]. Normally, NF-κB activation promotes the secretion of proinflammatory cytokines such as IL-12, which support Th1 responses. Disruption of NF-κB reduces IL-12, favoring Th2 differentiation. Additionally, impaired NF-κB signaling increases Th2 cytokines (IL-4, IL-5, and IL-13) crucial for Th2 responses. NF-κB is also essential for Treg function and its impairment leads to Treg dysfunction, reducing suppression of Th2 cells and promoting unchecked Th2 responses. Moreover, NF-κB regulates GATA3, a key Th2 differentiation factor, and its dysregulation further enhances Th2 polarization. These combined effects ultimately favor Th2 differentiation and allergic inflammation [29].

#### Abnormal T cell development and/or restriction of the T cell receptor repertoire

Limited TCR repertoire is also another mechanism that leads to Th2 skewing and allergic inflammation. Several genes involved in TCR repertoire selection and T cell development are implicated in PADs including *RAG1*, *RAG2*, *DCLRE1C*, *ADA*, *IL2RG*, *IL7RA*, *CHD7*, *LIG4*, *ZAP70*, *TBX1*, as well as 22q11.2 deletion syndrome. A powerful illustrative example is *RAG1* and *RAG2*, which are essential for V(D)J recombination and generating diverse TCRs necessary for a robust immune response [30]. Hypomorphic LOF variants in these *RAG* genes lead to restricted TCR diversity causing Omenn syndrome, characterized by Th2-skewed inflammation, eczema, elevated IgE, and eosinophilia [30]. The mechanism by which limited TCR repertoire can lead to atopy is not entirely clear. It has been previously postulated that reduced TCR diversity can create gaps in Treg repertoires, which limits their ability to control effector T cell responses [31,32]. Additionally, the absence of higher-affinity antigen-specific TCRs during naive T cell priming allows lower-affinity CD4<sup>+</sup> clones to expand noncompetitively and differentiate into Th2 effectors [33].

#### Altered cytokine signaling

Proallergic Th2 skewing can also be caused by variants that disrupt cytokine signaling. These damaging variants function through GOF, LOF, or DN mechanisms and to date have been found to cause PADs by disrupting each of the key steps in cytokine signaling, including cell-surface receptors (encoded by *IL2RA*, *IL4RA*, *TGFBR1*, *TGFBR2*, *IL6ST*, and *IL6R*), intracellular signaling molecules (encoded by *JAK1* and *ERBB2IP*), and transcription factors (encoded by *STAT1*, *STAT3*, *STAT5B*, *STAT6*, *ZNF341*, *FOXP3*, and *TBX21*). A recently described example is *STAT6*-GOF disease, which is caused by GOF variants in *STAT6* and results in enhanced IL-4 and IL-13 signaling through sustained *STAT6* phosphorylation and increased *STAT6* target gene expression [34,35]. This promotes Th2 cell differentiation and a skewing towards Th2 immune responses, amplifying allergic phenotypes such as atopic dermatitis, asthma, and elevated serum IgE levels. Additionally, many of these cytokine-related variants also impair Treg development and function, which is crucial for maintaining immune tolerance and preventing excessive allergic inflammation. Variants in *IL2RA*, *STAT5B*, and *FOXP3*, for instance, are well-documented to compromise Treg differentiation, survival, or suppressive capacity, thereby exacerbating Th2-driven inflammation [1,3]. Thus, Treg failure represents a key immunological consequence of cytokine signaling defects in PADs, further amplifying dysregulated immune responses.

### Altered cellular metabolism

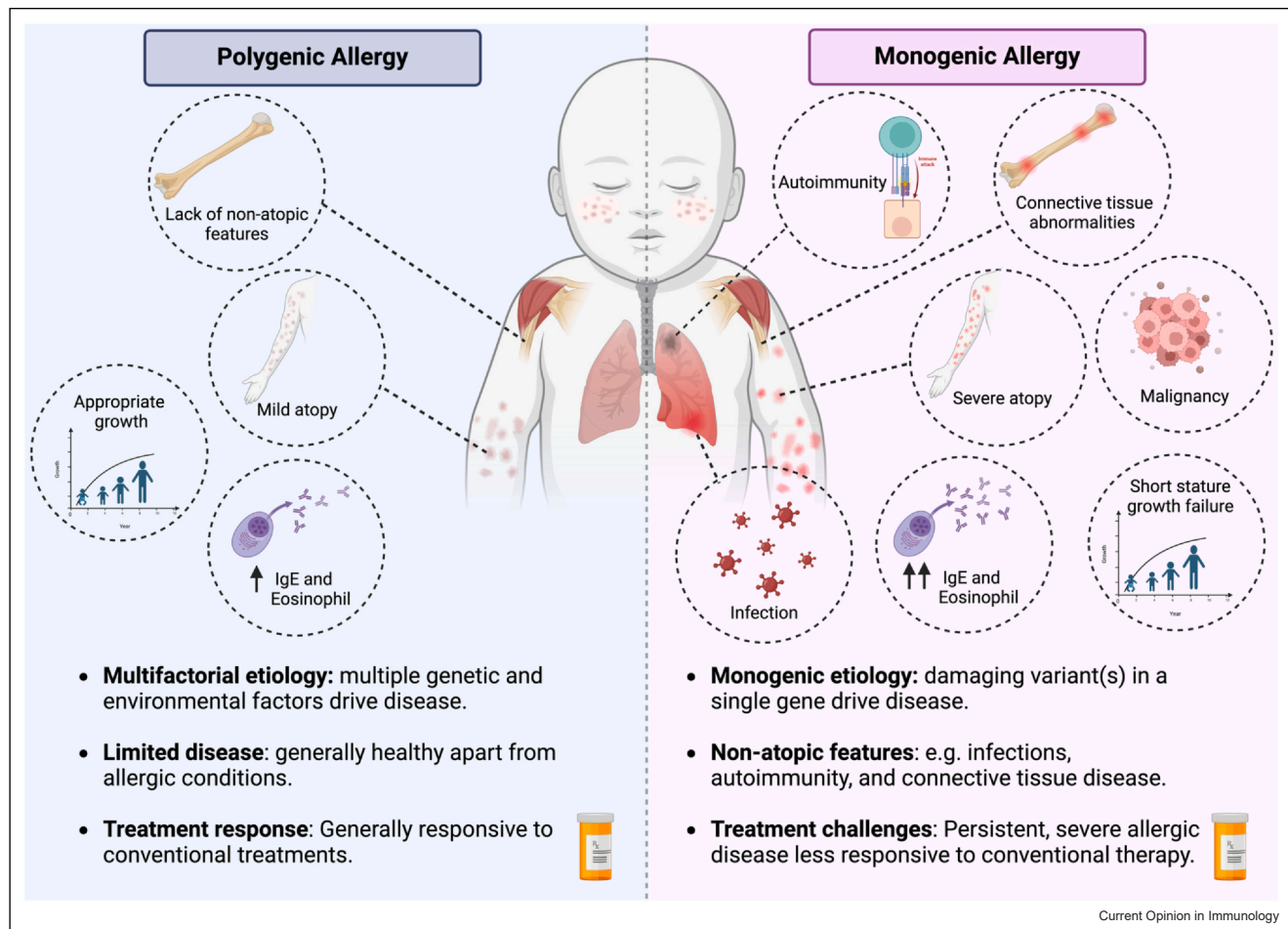
Finally, disruption of cellular metabolism is an emerging mechanism leading to the development of PADs. For example, LOF and DN variants in *CARD11* result in Th2 skewing through the disruption of the mechanistic target of rapamycin (mTOR) pathway, which is crucial for cell growth, proliferation, and metabolism [36]. Specifically, *CARD11* variants impair mTOR signaling by disrupting the expression of the amino acid transporter ASCT2, leading to decreased cellular glutamine uptake. This reduction in glutamine uptake impairs mTORC1 activation, subsequently reducing tricarboxylic acid cycle activity and glycolysis [36]. Th2 cells rely less on glycolysis than Th1 and Th17 cells; as such, disruptions in metabolic pathways that impact

glycolysis can preferentially support Th2 cell survival and proliferation, supporting a milieu conducive to allergic inflammation.

### Clinical features of primary atopic disorders

Although differentiating PADs from common polygenic allergic diseases poses a challenge for clinicians, the nature and severity of symptoms can provide a clue (Figure 1). Common allergic diseases typically present with isolated conditions such as atopic dermatitis, asthma, and allergic rhinitis, or may follow the classic childhood chronological progression referred to as the 'atopic march' that begins with atopic dermatitis [37]. Polygenic versions of atopic disease typically begin in early childhood but can develop at any age, often follow

Figure 1



Differentiating factors between polygenic and monogenic allergy. This figure illustrates the clinical and diagnostic distinctions between common polygenic allergic disease and monogenic allergic disease (also known as primary atopic disorders). Individuals with polygenic allergy generally present with a disease that is limited to allergic manifestations and that follows a typical childhood progression known as the 'atopic march'. Primary atopic disorders frequently involve additional nonatopic features, such as increased susceptibility to infections, autoimmunity, malignancy, growth impairment, and connective tissue disease. Atopic manifestations often develop early in life, are severe, and are resistant to conventional treatments. The figure was created using Biorender ([www.biorender.com](http://www.biorender.com)).

a waxing and waning disease course, and generally respond well to conventional therapies. In contrast, PADs manifest with more severe, persistent symptoms that typically emerge in the first months or year of life and often show an inadequate response to first-line therapies that are effective for common polygenic disease. Another key warning sign for PADs is the finding of additional nonatopic disease manifestations, such as increased susceptibility to infections, autoimmunity, and systemic features including failure to thrive, connective tissue abnormalities, and gastrointestinal inflammation [4].

Certain clinical features can provide insight into the specific PAD that may be underlying a patient's condition. For example, variants in genes responsible for skin barrier function, such as *DSG1*, *CDSN*, *DSP*, and *SPINK5*, result in conditions with severe skin involvement, including severe dermatitis, erythroderma, ichthyosis, and pruritus [5]. In contrast, connective tissue anomalies are a hallmark of TGF- $\beta$  pathway disorders, whether caused by damaging genetic changes in *STAT3* or by affecting key signaling components in the *STAT3* pathway such as *ZNF341*, *DOCK8*, *IL6ST*, *IL6R*, or *ERBIN*. However, PADs have many overlapping features making it very challenging to assign a probable gene or probable impacted pathway to individual patients in the clinical setting. For this reason, we strongly advocate for comprehensive gene sequencing using broadly inclusive gene panels, exome, or genome sequencing as the most efficient first-line strategy for diagnosing PADs.

### Diagnosis of primary atopic disorders

The classic clinical approach of history taking and physical examination, followed by the judicious use of laboratory evaluation, should be followed when seeing a patient with a possible PAD (Figure 1). In addition to the history of very early-onset severe disease, family history can be most informative. Common polygenic allergic diseases often show a family history of atopic conditions like asthma, atopic dermatitis, and allergic rhinitis without a clear pattern of inheritance. In contrast, PADs may follow a clear inheritance pattern such as autosomal dominant, autosomal recessive, or X-linked inheritance. However, it is important to note that spontaneously occurring *de novo* variants can also be a cause of PADs, so a PAD should not be dismissed in the absence of a family history or a clear Mendelian pattern of inheritance. History and physical examination should also focus on ruling out the possibility of underlying malignancy or infection as a possible explanation for the complex allergic presentation.

Laboratory tests are also often used in the diagnosis and workup of PADs and in distinguishing them from common allergic conditions. The biomarkers of allergic disease, namely peripheral blood eosinophil counts and serum IgE levels, are typically elevated in both polygenic and PADs.

However, it is not possible to define an IgE or eosinophil cutoff above which every patient must have a PADs workup, so clinical acumen is essential. In addition to IgE and eosinophil levels, more advanced diagnostic modalities to interrogate immune function may be informative, including serum immunoglobulin levels, assessing antibody production following immunization, flow cytometry to quantify cell types, as well as *in vitro* functional immune assays [38,39].

While symptoms, family history, and diagnostic laboratory tests provide essential insights, they are often not enough for definitively diagnosing PADs due to the variability in clinical presentation, the presence of overlapping symptoms with common allergic diseases, and the potential for atypical presentations. Genetic sequencing has become an essential tool in the diagnosis and management of PADs. Technologies such as targeted gene panels, exome sequencing, and genome sequencing should now be used early in the diagnostic workup to identify genetic variants responsible for these conditions. Over the past decade, next-generation sequencing (NGS) techniques have facilitated the evaluation of many patients, identifying both known and novel disease-causing genes. While there are no studies systematically defining the diagnostic yield of NGS for PADs, the results for monogenic IEIs are compelling. For example, an exome sequencing study of 303 IEI patients in Türkiye reported a 41.1% diagnostic yield, identifying 52 novel variants and underscoring the effectiveness of genomic approaches in uncovering new disease-associated genes [40]. However, diagnostic yield varies across cohorts and methodologies. A study by Posey et al. reported a 28.2% diagnostic rate for exome sequencing in a broader cohort of individuals with suspected genetic disorders, including IEIs [41], while Similuk et al. found 32.7% diagnostic yield specifically in US-based IEI cohorts [42]. These differences may reflect heterogeneity in patient selection, sequencing depth, and analytical pipelines. Despite variability in yield, NGS remains a powerful tool for genetic diagnosis in IEIs, particularly for patients with well-characterized PADs or those with suggestive clinical phenotypes. Beyond improving diagnostic rates, molecular testing has significant implications for clinical decision-making and patient outcomes. Molecularly defined diagnoses for patients with IEIs have been reported to alter therapeutic options in 34% of the 254 cases included in this study, significantly impacting patient care by enabling early diagnosis, personalized treatment strategies, and accurate genetic counseling and prognostic assessments [43].

### Discussion and conclusion

PADs are emerging as conditions that are more prevalent than one might imagine for conditions traditionally considered to be extremely rare. Although the prevalence of these diseases has not been formally assessed, PADs are part of the broader category of IEIs, which are

estimated to affect between 1 in 1000 and 1 in 5000 individuals globally [44]. Given this higher-than-expected prevalence, clinicians should be aware of the signs and appropriate workup and diagnostic methods for PADs, as accurate genetic diagnosis can have important implications for management and patient outcomes.

Genetic sequencing has revolutionized the diagnosis and management of PADs. Techniques such as targeted gene panels, exome sequencing, and genome sequencing have been effective in identifying both known and novel genetic variants responsible for these conditions [40]. Furthermore, molecularly defined diagnoses have significantly altered therapeutic options in a substantial proportion of cases [38,39]. Beyond their impact on patient management and outcomes, the discovery and characterization of PADs have significantly advanced our understanding of human immunology and the cellular and molecular mechanisms underlying allergic inflammation [4,45]. Furthermore, the study of the pathophysiology of these diseases has led to the development of precision therapeutics used to treat both monogenic and polygenic allergic diseases [5]. In fact, there are many examples in the clinical literature showing that molecular diagnosis has been a powerful tool in guiding targeted therapy, transforming outcomes for patients with PADs by precisely identifying the affected pathways [5]. For example, in JAK1 GOF disease, hyperactive JAK-STAT signaling drove severe atopy, eosinophilia, and hepatosplenomegaly — treatment with ruxolitinib (a JAK1/2 inhibitor) has led to dramatic clinical improvement in the affected individuals, including resolution of dermatitis, eosinophilia, and failure to thrive [46,47]. Another striking disease is STAT6-GOF disease, where aberrant IL-4/IL-13 signaling drives systemic allergic inflammation — targeting IL-4R $\alpha$  with dupilumab not only resolved eczema in the affected individuals but also significantly improved eosinophilia and growth [34,35]. Similarly, based upon the mechanistic demonstration of heightened IL-4R $\alpha$  expression in patients with ERBIN deficiency, molecularly informed treatment with dupilumab was highly effective in treating a patient with refractory severe atopy [48,49]. These clinical cases underscore how precise molecular diagnosis enables pathway-specific therapies, leading to transformative, personalized treatment approaches in rare immune disorders.

In conclusion, as genetic sequencing becomes more affordable, accessible, and commonly employed in the clinical setting, the number of diagnosed cases of PADs will rise. Maintaining a high index of suspicion for PADs and utilizing advanced diagnostic tools are crucial for the accurate and timely diagnosis of the conditions, leading to improved patient outcomes.

## Funding

This work was supported by grants from the Canadian Institutes of Health Research (PJO-173584 to S.E.T. and C.M.B.) and Genome British Columbia (SIP007) (S.E.T.). S.E.T. holds a Tier 1 Canada Research Chair in Pediatric Precision Health and the Aubrey J. Tingle Professor of Pediatric Immunology. C.M.B. is the recipient of a Health Professional-Investigator award from Michael Smith Health Research BC. M.V.S. is funded by the Vanier Canada Graduate Scholarship (Vanier CGS) and the University of British Columbia Four Year Doctoral Fellowship (4YF). S.S. is funded by the University of British Columbia Four Year Doctoral Fellowship (4YF).

## Data Availability

No data were used for the research described in the article.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

•• of outstanding interest

1. Lyons JJ, Milner JD: **Primary atopic disorders**. *J Exp Med* 2018, **215**:1009-1022.
2. Lyons JJ, Milner JD: **The clinical and mechanistic intersection of primary atopic disorders and inborn errors of growth and metabolism**. *Immunol Rev* 2019, **287**:135-144.
3. Milner JD: **Primary atopic disorders**. *Annu Rev Immunol* 2020, **38**:785-808.
4. Vaseghi-Shanjani M, Smith KL, Sara RJ, Modi BP, Branch A, Sharma M, Lu HY, James EL, Hildebrand KJ, Biggs CM, et al.: **Inborn errors of immunity manifesting as atopic disorders**. *J Allergy Clin Immunol* 2021, **148**:1130-1139.
5. Vaseghi-Shanjani M, Snow AL, Margolis DJ, Latrous M, Milner JD, Turvey SE, Biggs CM: **Atopy as immune dysregulation: offender genes and targets**. *J Allergy Clin Immunol Pract* 2022, **10**:1737-1756.
6. Nelson RW, Geha RS, McDonald DR: **Inborn errors of the immune system associated with atopy**. *Front Immunol* 2022, **13**:860821.
7. Yang R, Mele F, Worley L, Langlais D, Rosain J, Benhsaïen I, Elarabi H, Croft CA, Doisne J-M, Zhang P, et al.: **Human T-bet governs**

- innate and innate-like adaptive IFN- $\gamma$ ; immunity against mycobacteria. *Cell* 2020, **183**:1826-1847.e1831.
8. Yang R, Weisshaar M, Mele F, Benhsaien I, Dorgham K, Han J, Croft CA, Notarbartolo S, Rosain J, Bastard P, et al.: **High Th2 cytokine levels and upper airway inflammation in human inherited T-bet deficiency.** *J Exp Med* 2021, **218**:e20202726.
  9. James AE, Abdalgani M, Khoury P, Freeman AF, Milner JD: **T(H)2-driven manifestations of inborn errors of immunity.** *J Allergy Clin Immunol* 2024, **154**:245-254.
- This review by Alyssa E. James et al. explores the impact of Th2-driven immune responses in IEI. The authors identify specific genetic variants that lead to a range of clinical manifestations, including autoimmunity, immunodeficiency, and allergic inflammation, particularly in cases of early-onset or atypical allergic diseases. They emphasize the critical role of genetic sequencing for accurate diagnosis and management, suggesting that targeted therapies based on genetic findings could enhance patient outcomes. This study is significant as it advances the understanding of IEI pathophysiology and highlights the need for integrating genetic testing into clinical practice to improve treatment strategies for affected individuals.
10. van den Bogaard EH, Elias PM, Goleva E, Berdyshev E, Smits JPH, Danby SG, Cork MJ, Leung DYM: **Targeting skin barrier function in atopic dermatitis.** *J Allergy Clin Immunol Pract* 2023, **11**:1335-1346.
  11. Wang J, Zhou Y, Zhang H, Hu L, Liu J, Wang L, Wang T, Zhang H, Cong L, Wang Q: **Pathogenesis of allergic diseases and implications for therapeutic interventions.** *Signal Transduct Target Ther* 2023, **8**:138.
  12. Virolainen SJ, Satish L, Biagini JM, Chaib H, Chang WC, Dexheimer PJ, Dixon MR, Dunn K, Fletcher D, Forney C, et al.: **Filaggrin loss-of-function variants are associated with atopic dermatitis phenotypes in a diverse, early-life prospective cohort.** *JCI Insight* 2024, **9**.
  13. Lu HF, Zhou YC, Yang LT, Zhou Q, Wang XJ, Qiu SQ, Cheng BH, Zeng XH: **Involvement and repair of epithelial barrier dysfunction in allergic diseases.** *Front Immunol* 2024, **15**:1348272.
  14. Laky K, Kinard JL, Li JM, Moore IN, Lack J, Fischer ER, Kabat J, Latanich R, Zachos NC, Limkar AR, et al.: **Epithelial-intrinsic defects in TGF $\beta$  signaling drive local allergic inflammation manifesting as eosinophilic esophagitis.** *Sci Immunol* 2023, **8**:eabp9940.
  15. Radtke D, Voehringer D: **Granulocyte development, tissue recruitment, and function during allergic inflammation.** *Eur J Immunol* 2023, **53**:e2249977.
  16. Garcia-Montero AC: **KIT mutation in mast cells and other bone marrow hematopoietic cell lineages in systemic mast cell disorders: a prospective study of the Spanish Network on Mastocytosis (REMA) in a series of 113 patients.** *Blood* 2006, **108**:2366-2372.
  17. Ombrello MJ, Remmers EF, Sun G, Freeman AF, Datta S, Torabi-Parizi P, Subramanian N, Bunney TD, Baxendale RW, Martins MS, et al.: **Cold urticaria, immunodeficiency, and autoimmunity related to PLCG2 deletions.** *N Engl J Med* 2012, **366**:330-338.
  18. Boyden SE, Desai A, Cruse G, Young ML, Bolan HC, Scott LM, Eisch AR, Long RD, Lee C-CR, Satorius CL, et al.: **Vibratory urticaria associated with a missense variant in ADGRE2.** *N Engl J Med* 2016, **374**:656-663.
  19. Tangye SG, Bucciol G, Casas-Martin J, Pillay B, Ma CS, Moens L, Meyts I: **Human inborn errors of the actin cytoskeleton affecting immunity: way beyond WAS and WIP.** *Immunol Cell Biol* 2019, **97**:389-402.
  20. Vasquez-Echeverri E, Yamazaki-Nakashimada MA, Venegas Montoya E, Scheffler Mendoza SC, Castano-Jaramillo LM, Medina-Torres EA, Gonzalez-Serrano ME, Espinosa-Navarro M, Bustamante Ogando JC, Gonzalez-Villarreal MG, et al.: **Is your kid actin out? A series of six patients with inherited actin-related protein 2/3 complex subunit 1B deficiency and review of the literature.** *J Allergy Clin Immunol Pract* 2023, **11**:1261-1280 e1268.
  21. Fang Y, Luo Y, Liu Y, Chen J: **A novel variant of X-linked moesin gene in a boy with inflammatory bowel disease like disease — a case report.** *Front Genet* 2022, **13**.
  22. Cagdas D, Halacli SO, Tan C, Esenboga S, Karaatmaca B, Cetinkaya PG, Balci-Hayta B, Ayhan A, Uner A, Orhan D, et al.: **Diversity in serine/threonine protein kinase-4 deficiency and review of the literature.** *J Allergy Clin Immunol Pract* 2021, **9**:3752-3766 e3754.
  23. Alazami AM, Al-Helale M, Alhissi S, Al-Saud B, Alajlan H, Monies D, Shah Z, Abouelhoda M, Amaout R, Al-Dhekri H, et al.: **Novel CARMIL2 mutations in patients with variable clinical dermatitis, infections, and combined immunodeficiency.** *Front Immunol* 2018, **9**.
  24. Cook SA, Comrie WA, Poli MC, Similuk M, Oler AJ, Faruqi AJ, Kuhns DB, Yang S, Vargas-Hernandez A, Carisey AF, et al.: **HEM1 deficiency disrupts mTORC2 and F-actin control in inherited immunodysregulatory disease.** *Science* 2020, **369**:202-207.
  25. Tangye SG, Pillay B, Randall KL, Avery DT, Phan TG, Gray P, Ziegler JB, Smart JM, Peake J, Arkwright PD, et al.: **Dedicator of cytokinesis 8-deficient CD4+ T cells are biased to a TH2 effector fate at the expense of TH1 and TH17 cells.** *J Allergy Clin Immunol* 2017, **139**:933-949.
  26. Alroqi FJ, Charbonnier L-M, Keles S, Ghandour F, Mouawad P, Sabouneh R, Mohammed R, Almutairi A, Chou J, Massaad MJ, et al.: **DOCK8 deficiency presenting as an IPEX-like disorder.** *J Clin Immunol* 2017, **37**:811-819.
  27. van Panhuys N, Klauschen F, Germain RN: **T-cell-receptor-dependent signal intensity dominantly controls CD4(+) T cell polarization In Vivo.** *Immunity* 2014, **41**:63-74.
  28. Ouederni M, Vincent QB, Frange P, Touzot F, Scerra S, Bejaoui M, Bousfiha A, Levy Y, Lisowska-Grosj Pierre B, Canioni D, et al.: **Major histocompatibility complex class II expression deficiency caused by a RFXANK founder mutation: a survey of 35 patients.** *Blood* 2011, **118**:5108-5118.
  29. Oh H, Ghosh S: **NF- $\kappa$ B: roles and regulation in different CD4+ T-cell subsets.** *Immunol Rev* 2013, **252**:41-51.
  30. Braams M, Pike-Overzet K, Staal FJT: **The recombinase activating genes: architects of immune diversity during lymphocyte development.** *Front Immunol* 2023, **14**:1210818.
  31. Milner JD, Ward JM, Keane-Myers A, Paul WE: **Lymphopenic mice reconstituted with limited repertoire T cells develop severe, multiorgan, Th2-associated inflammatory disease.** *Proc Natl Acad Sci* 2007, **104**:576-581.
  32. Lawrence MG, Barber JS, Sokolic RA, Garabedian EK, Desai AN, O'Brien M, Jones N, Bali P, Hershfield MS, Stone KD, et al.: **Elevated IgE and atopy in patients treated for early-onset ADA-SCID.** *J Allergy Clin Immunol* 2013, **132**:1444-1446.e1445.
  33. Milner JD, Fazilleau N, McHeyzer-Williams M, Paul W: **Cutting edge: lack of high affinity competition for peptide in polyclonal CD4+ responses unmasks IL-4 production.** *J Immunol* 2010, **184**:6569-6573.
  34. Sharma M, Leung D, Momenilandi M, Jones LCW, Pacillo L, James AE, Murrell JR, Delafontaine S, Maimaris J, Vaseghi-Shanjani M, et al.: **Human germline heterozygous gain-of-function STAT6 variants cause severe allergic disease.** *J Exp Med* 2023, **220**.
- This study describes the assembly of a global consortium to identify STAT6 GOF disease in 16 patients from 10 families spanning three continents. Patients all presented with a profound phenotype of early-life onset allergic immune dysregulation, widespread treatment-resistant atopic dermatitis, hypereosinophilia with eosinophilic gastrointestinal disease, asthma, elevated serum IgE, IgE-mediated food allergies, and anaphylaxis. All patients carried monoallelic rare variants in STAT6 and mechanistic studies established their GOF phenotype. Treatment with the anti-IL-4R $\alpha$  antibody, dupilumab, was highly effective.
35. Sharma M, Suratannon N, Leung D, Baris S, Takeuchi I, Samra S, Yanagi K, Rosa Duque J: **Human germline gain-of-function in STAT6: from severe allergic disease to lymphoma and beyond.** *Trends Immunol* 2024, **45**:138-153.
  36. Ma CA, Stinson JR, Zhang Y, Abbott JK, Weinreich MA, Hauk PJ, Reynolds PR, Lyons JJ, Nelson CG, Ruffo E, et al.: **Germline**

**hypomorphic CARD11 mutations in severe atopic disease.** *Nat Genet* 2017, **49**:1192-1201.

37. Paller AS, Spergel JM, Mina-Orsorio P, Irvine AD: **The atopic march and atopic multimorbidity: many trajectories, many pathways.** *J Allergy Clin Immunol* 2019, **143**:46-55.
  38. Bucciol G, Van Nieuwenhove E, Moens L, Itan Y, Meyts I: **Whole exome sequencing in inborn errors of immunity: use the power but mind the limits.** *Curr Opin Allergy Clin Immunol* 2017, **17**:421-430.
  39. Conley ME, Casanova J-L: **Discovery of single-gene inborn errors of immunity by next generation sequencing.** *Curr Opin Immunol* 2014, **30**:17-23.
  40. Erman B, Aba U, Ipsir C, Pehlivan D, Aytakin C, Cildir G, Cicek B, Bozkurt C, Tekeoglu S, Kaya M, et al.: **Genetic evaluation of the patients with clinically diagnosed inborn errors of immunity by whole exome sequencing: results from a specialized research center for immunodeficiency in Türkiye.** *J Clin Immunol* 2024, **44**.
- This study evaluated 303 patients with clinically diagnosed IEI using exome sequencing (ES) at a specialized research center in Türkiye. The genetic analysis resulted in a diagnosis for 41.1% of the patients, identifying 127 variants, 52 of which were novel. The diagnostic yield varied significantly among different IEI subtypes, with a 100% diagnostic rate for Severe Combined Immunodeficiency and lower rates for conditions like Primary Antibody Deficiency at 15.6%. The findings underscore the importance of ES in identifying genetic causes of IEI, particularly in consanguineous populations, and highlight the potential of ES to improve the diagnosis and treatment of these complex disorders.
41. Posey JE, Harel T, Liu P, Rosenfeld JA, James RA, Coban Akdemir ZH, Walkiewicz M, Bi W, Xiao R, Ding Y, et al.: **Resolution of disease phenotypes resulting from multilocus genomic variation.** *N Engl J Med* 2017, **376**:21-31.
  42. Similuk MN, Yan J, Ghosh R, Oler AJ, Franco LM, Setzer MR, Kamen M, Jodarski C, Dimaggio T, Davis J, et al.: **Clinical exome sequencing of 1000 families with complex immune phenotypes:**

**toward comprehensive genomic evaluations.** *J Allergy Clin Immunol* 2022, **150**:947-954.

43. Arts P, Simons A, Alzahrani MS, Yilmaz E, Alidrisi E, Van Aerde KJ, Alenezi N, Alghamdi HA, Aljubab HA, Al-Hussaini AA, et al.: **Exome sequencing in routine diagnostics: a generic test for 254 patients with primary immunodeficiencies.** *Genome Med* 2019, **11**.
44. Tangye SG, Al-Herz W, Bousfiha A, Cunningham-Rundles C, Franco JL, Holland SM, Klein C, Morio T, Oksenhendler E, Picard C, et al.: **Human inborn errors of immunity: 2022 update on the classification from the International Union of Immunological Societies Expert Committee.** *J Clin Immunol* 2022, **42**:1473-1507.
45. Vaseghi-Shanjani M, Yousefi P, Sharma M, Samra S, Sifuentes E, Turvey SE, Biggs CM: **Transcription factor defects in inborn errors of immunity with atopy.** *Front Allergy* 2023, **4**:1237852.
46. Del Bel KL, Ragotte RJ, Saferali A, Lee S, Vercauteren SM, Mostafavi SA, Schreiber RA, Prendiville JS, Phang MS, Halperin J, et al.: **JAK1 gain-of-function causes an autosomal dominant immune dysregulatory and hypereosinophilic syndrome.** *J Allergy Clin Immunol* 2017, .
47. Biggs CM, Cordeiro-Santanach A, Prykhodzhiy SV, Deveau AP, Lin Y, Del Bel KL, Orben F, Ragotte RJ, Saferali A, Mostafavi S, et al.: **Human JAK1 gain of function causes dysregulated myelopoeisis and severe allergic inflammation.** *JCI Insight* 2022, **7**.
48. Lyons JJ, Liu Y, Ma CA, Yu X, O'Connell MP, Lawrence MG, Zhang Y, Karpe K, Zhao M, Siegel AM, et al.: **ERBIN deficiency links STAT3 and TGF- $\beta$  pathway defects with atopy in humans.** *J Exp Med* 2017, **214**:669-680.
49. Droghini HR, Abonia JP, Collins MH, Milner JD, Lyons JJ, Freeman AF, Mukkada VA, Risma KA, Rothenberg ME, Schwartz JT: **Targeted IL-4R $\alpha$  blockade ameliorates refractory allergic eosinophilic inflammation in a patient with dysregulated TGF- $\beta$  signaling due to ERBIN deficiency.** *J Allergy Clin Immunol Pract* 2022, **10**:1903-1906.