

# **Respiratory infections in lung transplant recipients**

Christopher Alexander Hinze<sup>a,b</sup>, Susanne Simon<sup>a</sup> and Jens Gottlieb<sup>a,b</sup>

#### **Purpose of review**

Morbidity and mortality rates after lung transplantation still remain higher than after other forms of solid organ transplantation, primarily due to a higher risk of infections and the development of chronic lung allograft dysfunction. Thus, a tiered approach highlighting the most significant respiratory pathogens including common opportunistic infections along with diagnostic, treatment and prevention strategies, including vaccination and prophylaxis is needed.

#### **Recent findings**

The need for intense immunosuppressive therapy to prevent rejection, coupled with the transplanted lung's constant exposure to environment and impaired local defence mechanisms leads to frequent infections. Viral and bacterial infections are most frequent while fungal infections mainly involve the tracheobronchial tract but may be fatal in case of disseminated disease. Some infectious agents are known to trigger acute rejection or contribute to chronic allograft dysfunction. Invasive testing in the form of bronchoscopy with bronchoalveolar lavage is standard and increasing experience in point of care testing is gained to allow early preemptive therapy.

#### Summary

Timely diagnosis, treatment, and ongoing monitoring are essential, but this can be difficult due to the wide variety of potential pathogens.

#### Keywords

community-acquired respiratory viruses, chronic lung allograft dysfunction, lung transplantation, respiratory infection

#### INTRODUCTION

Respiratory infections are a major complication after lung transplantation (LTx). They are associated with a high burden of disease and potentially life threatening either through immediate graft damage and/ or its association with the development of chronic lung allograft dysfunction (CLAD). Besides immunosuppression, susceptibility to respiratory infections is increased because of impaired mucociliary clearance and dampened cough reflex due to organ denervation, potential colonization of the graft and continued exposure to the environment [1–3]. Overall infections are the most prevalent adverse events following immunosuppressive drugs. Of those bacterial infections represent the majority, followed by viral and fungal infections during a 3-year observational period [4<sup>•••</sup>]. The spectrum of respiratory pathogens is significantly broader in lung transplant patients than in other solid organ recipients. In addition to known pathogens, pathogens such as Nocardia, nontuberculous mycobacteria (NTM), fungal infections (especially Aspergillus spp.), CMV, Pneumocystis jirovecii and a broad spectrum of respiratory viruses should also be included if a respiratory infection is suspected [5]. During the first month after transplantation, infections are primarily bacterial, including hospital-acquired, donor- and recipient-related or affecting the bronchial anastomosis. Afterwards, viral reactivations and opportunistic infections are common, while community-acquired pathogens become increasingly significant in late-onset infections [1,6].

## VIRAL INFECTIONS

## Community acquired respiratory viruses

Community acquired respiratory viruses (CARV) are a common threat in lung transplant recipients. Data

<sup>a</sup>Department of Respiratory Medicine and Infectious Diseases, Hannover Medical School and <sup>b</sup>Biomedical Research in Endstage and Obstructive Lung Disease Hannover (BREATH), Member of the German Center for Lung Research (DZL), Hannover, Germany

Correspondence to Christopher Alexander Hinze, MD, Department of Respiratory Medicine and Infectious Disease, Hannover Medical School, Hannover, Germany; Carl-Neuberg-Str. 1, 30625 Hannover, Germany. Tel: +49 511 532 3531; fax: +49 511 532 1119; e-mail: hinze.christopher@mh-hannover.de

Curr Opin Infect Dis 2025, 38:150–160

DOI:10.1097/QCO.000000000001097

www.co-infectiousdiseases.com

Volume 38 • Number 2 • April 2025

# **KEY POINTS**

- Respiratory infections following lung transplantation carry significant risks, including immediate graft damage and chronic lung allograft dysfunction (CLAD).
- Community-acquired respiratory viruses (CARV) often cause serious complications in lung transplant recipients, including prolonged cough and graft dysfunction.
- Diagnostic approaches, including PCR, bronchoalveolar lavage, and point-of-care testing, are essential for rapid and accurate identification of infections to initiate timely treatment.
- Preventive strategies such as vaccination, lifelong prophylactic drug therapy and early antiviral treatment aim to reduce infection-related risks and improve survival for lung transplant patients.

about infection incidence vary between 10 and 68 cases/100 patient years among CARV [7]. Subtypes include influenza A/B, SARS-CoV-2, seasonal coronaviruses, human metapneumovirus-hMPV, respiratory syncytial virus (RSV), parainfluenza, rhinovirus, enterovirus and adenovirus [8-11]. Seasonal occurrence in the period between October and April is observed except for adenovirus, rhinovirus and parainfluenza virus [11-13]. Viral infections after lung transplantation not only cause potentially life-threatening pneumonia with higher risk for bacterial and fungal superinfection, but are also associated with an increased incidence or progression of chronic lung allograft dysfunction (CLAD). CLAD is a difficult to treat complication after LTx and is associated with increased mortality [12,14,15].

# **Clinical presentation and diagnostic**

Symptoms and disease severity differ considerably between patients. CARV infection may cause symptoms of upper respiratory tract infection (URTI) e.g. sore throat, rhinorrhea or lower respiratory tract infections (LRTI) e.g. cough, wheezing and graft dysfunction assessed by declining forced expiratory volume (FEV<sub>1</sub>) and even signs of respiratory failure [16,17]. In such cases, a diagnostic workup consisting of clinical, radiological and laboratory tests should be performed (Fig. 1). Chest radiographic and computer tomography scans (CT) may reveal heterogeneous abnormalities including ground glass opacities and consolidations [18,19].

For diagnosis there are several multiplex PCR kits commercially available for nasopharyngeal swabs as well as for samples from the lower

respiratory tract including bronchoalveolar lavage [20]. Upper and lower respiratory tract sampling are frequently used as diagnostic tools in daily clinical practice. Although bronchoscopy implicates invasiveness and potential periinterventional complications it represents a safe procedure even in the outpatient setting after LTx [21]. A high concordance between both sampling methods was detected depending on viral pathogen. Bronchoalveolar lavage provides additional information especially regarding bacterial and fungal co-infections and noninfectious cases of graft dysfunction [22-24.25<sup>•</sup>] (Fig. 1). The emergence of point-of-care tests (POCT) offers the opportunity to significantly accelerate the diagnosis of viral respiratory tract infections to rapidly initiate specific therapy. The diagnostic power is slightly lower [26<sup>•</sup>,27<sup>•</sup>] but return time for results is shorter usually by one day. When using POCT patient profiles should be defined to identify those who might benefit from POCT, for example, patients at high risk for severe disease.

Bronchoscopy is considered the gold standard for diagnosis of infections after LTx. Pathogen isolation is expected to be successful in 60% of LTx patients with infection, 20% will have a noninfectious cause mimicking infection and 20% of the cases remain unclear even after bronchoscopic work-up.

Infection with SARS-CoV-2 still represents a severe complication for lung transplant patients and is still associated with an elevated mortality around 1-5% and severe course in up to 10% in the Omicron era [28",29"]. In the early phase of the pandemic mortality was reported to be up to 30% after LTx [30]. Thus, it is crucial to provide early therapy as well as identifying patients at risk of severe disease progression (age >60 years, glomerular filtration rate below 30 ml/min/1.73 m<sup>2</sup>). Among solid organ transplanted patients lung transplant recipients are most endangered [31,32"].

# Treatment

Despite advances during the last years therapeutic options for the treatment of CARV infections treatment is still challenging. Oseltamivir, an oral neuraminidase inhibitor is the main therapeutic option for lung transplant recipients to treat influenza A and B infection. Several studies have provided evidence that early therapy is associated with a reduction in mortality, ICU stays and severe pneumonia with no significant interactions and good tolerability [33–35]. Further drugs against influenza, for example, baloxavir (endonuclease inhibitor), Peramivir or Zanamivir (neuraminidase inhibitor) are



**FIGURE 1.** Diagnostic algorithm for lung transplanted patients with suspicion for lower respiratory tract infection. Ag, antigen; BAL, bronchoalveolar lavage; CARV, community acquired respiratory viruses; CLAD, chronic lung allograft dysfunction; CMV, cytomegalovirus; CNI, calcineurin-Inhibitor; CT, computer tomography; HLA, human leukocyte antigen; IGRA, interferon-gamma release assay; NTproBNP, N-terminal pro-brain natriuretic peptide; PCP, *Pneumocystis jirovecii*; PCR, polymerase chain reaction; POCT, point-of-care test; TBB, transbronchial biopsy; TBC, *Mycobacterium tuberculosis*.

152 www.co-infectiousdiseases.com



**FIGURE 2.** Therapeutic algorithm for Sars-CoV-2 infection after lung transplantation. (a) At least three antigen contacts consisting of at least two vaccines; ALT, alanin-aminotransferase; AST, aspartat-aminotransferase; BLI, betalactamase-inhibitor; CI, contraindication; CNI, calcineurin inhibitor; GFR, glomerulus filtration rate; HFNC, high flow nasal cannula; iv, intravenous; LTOT, long-term oxygen therapy; MMF, mycophenolate mofetil; mTOR, mechanistic target of rapamycin; MV, mechanical ventilation; NIV, noninvasive ventilation.

not approved for lung transplanted patients in all countries.

Cohort studies are pointing towards the direction that early therapy of intravenous Remdesivir is less frequently associated with progression to severe COVID-19 disease [31,32<sup>•</sup>] (Fig. 2). Nirmatrelvir/ ritonavir as an orally available drug is limited in use due to serious interactions with calcineurin inhibitors (CNI). There is no proven benefit and a high risk of CNI toxicity. If used close monitoring and extensive dose adjustments is needed [36].

Ribavirin is an antiviral agent against RSV and paramyxoviruses that may shorten lung function recovery after both RSV and paramyxoviruses and prevents development of bronchiolitis obliterans syndrome [12,37–39]. Oral administration appears to be more cost-effective and not inferior to intravenous administration due to the possibility of outpatient administration [40]. Unfortunately, there is no randomized controlled trial of ribavirin in paramyxoviruses to support its use. Other agents (Presatovir, ALN RSV01) have been studied in RCTs without proven benefit in RSV infected LTx recipients [41,42].

# Cytomegalovirus

Cytomegalovirus (CMV) belongs to herpes virus family that can reactivate or infect primarily people with impaired immune systems. Since the lungs are a key reservoir for CMV, lung transplantation tends to involve a higher transfer of CMV compared to other organ transplants [43,44]. A diagnosis of CMV pneumonitis requires histopathological confirmation of inclusion bodies and is usually accompanied by CMV-DNemia. A Positive PCR in BAL alone is not sufficient for diagnosis. CMV pneumonitis is linked to a significant rise in mortality and morbidity whereas already reduced CMV replication within the lung is associated with CLAD [45,46]. Although CMV pneumonia is relatively uncommon because of preemptive therapy, CMV-infection is diagnosed by CMV-DNAema before organ manifestation occurs. Additional diagnostic criteria including imaging results and a high CMV load in bronchoalveolar lavage supports the diagnosis [47,48].

Therapy involves reduction of immunosuppression (usually cell cycle inhibitors) and antiviral treatment. Mild disease with low levels of DNemia may be treated with oral valgancyclovir, while severe cases require intravenous ganciclovir until symptoms and viremia resolve. In case of refractory disease, ganciclovir resistance or intolerance of ganciclovir, alternative treatments like foscarnet, maribavir, or adoptive CMV-specific T cell therapy may be considered [49].

## **Bacterial infections**

Bacterial infections cause a high proportion of especially nosocomial complications early after transplantation, but are also a risk for the patient throughout the entire period after transplantation [50]. Tracheobronchitis must be differentiated from pneumonia with the latter being associated with severe disease and a higher incidence of respiratory failure [51]. Bacterial pathogens are responsible for 62–82% of cases of pneumonia in lung transplant recipients, most frequently caused by gram-negative species, especially Pseudomonas aeruginosa [50,52–54]. During ongoing follow-up after LTx, community-acquired pathogens like Streptococcus pneumoniae and Haemophilus influenzae gain importance. Pneumonia after LTx is associated with a risk of hospitalization, graft lost and death after LTx [55–57]. Patients become susceptible to these pathogens with increasing time after lung transplantation with considerably higher incidence than in the general population [56]. In general, LTx patients have more severe course of pneumonia and are frequently in need for prolonged treatment.

# **Opportunistic pathogens**

# Nocardia

Nocardia species are common environmental, opportunistic gram-positive bacteria that can cause both localized and widespread infections. Of all organ transplant recipients, lung transplants have the highest risk of infection affecting up to 3.5% of patients [58–60]. Risk factors include more intense steroids, advanced patient age and extended stays in the intensive care unit following transplantation [61]. Since the respiratory tract is the primary route of acquisition, the lungs are typically the first site of infection. Nocardiosis should be considered in LTx recipients who present with pneumonia, lung cavities or consolidations as well as pleural effusions visible on CT scans [1,62] (Fig. 3). If nocardiosis is diagnosed, brain imaging is recommended due to frequent disseminated (12.5-40% of cases), potentially asymptomatic disease to the central nervous system [58,61,62]. While surgical drainage may be required, antibiotic therapy remains the primary treatment for nocardiosis. First-line treatments typically involve carbapenems and cotrimoxazole. Alternative antibiotic regimens may be necessary if there is drug intolerance or resistance. After a prolonged intravenous therapy for several weeks oral therapy up to several months is usually initiated [63]. If the brain is infected, third-generation cephalosporins can be used due to good CNS access [63].

# Mycobacteria

Non tuberculous mycobacteria (NTM) occur ubiquitously in the environment and cause opportunistic infections, mainly in the lung and pleura. Lung transplanted patients are at high risk of developing NTM disease due to the chronic lung damage, high immunosuppression and continuous environmental exposure [64]. Furthermore, patients with cystic fibrosis are infected before transplanted and NTM may harbor in the upper respiratory tract [65]. According to IDSA/ATS guidelines, the fulfillment of clinical, radiological and microbiological criteria is needed to establish a diagnosis of pulmonary NTM [64,66]. Among the many known subspecies, Mycobacterium avium complex, Mycobacterium



**FIGURE 3.** Nocardiosis in 66 years old male 10 month after transplantation. Upper CT-chest with new onset right mediastinal mass (upper right) within 6 weeks between scans (upper left and right). Lower left: culture plate with white colonies, typical of nocardia.

abscessus and Mycobacterium gordonae are most frequently detected in lung transplant patients. Although overall survival is not affected by NTM infection, morbidity is high and side effects of treatment relevant [67,68]. Depending on the species, treatment consists of a combination of usually three to four antibiotics with different mechanisms of action for usually more than 12 months. The use of rifampicin leads to severe interactions with CNI, which necessitates close monitoring of levels and dose adjustment [69].

## **FUNGAL INFECTIONS**

## Pneumocystis jirovecii

*Pneumocystis jirovecii* (Pj) is an opportunistic fungal pathogen leading to rapidly progressing pneumonia, which may result in respiratory failure and death (up to 23% 90 days after infection) in transplanted individuals, especially lung transplant

recipients without prophylaxis [70-72]. Following the introduction of prophylaxis with sulfamethoxazole-trimethoprim, the infection rates among lung transplant recipients are rare [71,73]. Besides immunosuppression, increasing age, coincident CMVinfection and low total lymphocyte count were associated with a higher risk for severe Pj infection [73,74]. If symptoms like fever, cough and hypoxemia are present in a LTx patient without prophylaxis, Pj pneumonia (PjP) should be considered as a possible diagnosis. CT scans provide a higher sensitivity than a conventional X-ray chest [73,75]. Diagnosis of PjP relies on the direct detection of the microorganism in respiratory samples. PCR has become an increasingly valuable diagnostic tool due to its higher sensitivity compared to traditional microscopic methods [76]. Additionally, serum [1,3]-D-glucan as a component of the fungal cell wall, offers high sensitivity and specificity for Pj infection, serving as a useful complementary diagnostic tool [77,78]. The first-line treatment for Pj is

high-dose intravenous sulfamethoxazole-trimethoprim and in patients with a high suspicion of the disease, treatment should begin without delay, even before diagnostic results are confirmed [73,79].

## Aspergillus spp.

Infection with Aspergillus species impose serious threat for lung transplant recipients especially since they are ubiquitous in the environment and the respiratory tract is a major route of infection [80]. Aspergillus fumigatus accounted for the largest proportion of the pathogens detected [81,82]. Most centers worldwide are using universal prophylaxis with azoles or inhaled liposomal amphotericin B early after LTx [83]. Most aspergillus infections involve the tracheobronchial tree especially anastomoses and postanastomotic regions. Foreign bodies like stents and lose suture material are risk factors for chronic infection. This emphasizes the need for frequent endoscopic controls [84]. Disseminated disease is infrequent due to fungal prophylaxis; patients often suffer from systemic symptoms like fever and respiratory symptoms like cough, dyspnea, hemoptysis and in case of invasive growth chest/pleura pain [80]. In chest computer tomography, typical signs of invasive pulmonary aspergillosis include halo sign and macronodules [85]. Another manifestation of pulmonary aspergillosis is aspergilloma in preformed cavities in damaged transplant lungs (e.g. by CLAD) which is often accompanied by hemoptysis [86] (Fig. 4). Treatment of Aspergillus tracheobronchitis and anastomosis

infection consists of topical antifungals (e.g. inhaled liposomal amphotericin B) and oral broad spectrum azole therapy in conjunction with endoscopic debridement of necrotic tissue and removal of foreign bodies. In case of severe tracheobronchitis accompanied by necrosis or in case of untreatable aspergilloma, surgical interventions need to be considered [80,86,87]. In general prognosis of tracheobronchial infections after LTx is excellent in contrast to disseminated disease. Prolonged use of voriconazole in LTx patients is associated with an increased risk of skin cancer and should be avoided [88].

## **PREVENTIVE STRATEGIES**

# Vaccination

For organ transplant recipients, there is limited information about the response to vaccinations against respiratory pathogens. Limited humoral responses and a higher rate of infections after vaccination have been reported in LTx patients. No measures to improve vaccination response are established [56]. As the vaccination rates of the recommended vaccinations vary greatly, even after transplantation, it is essential to update all vaccinations before transplantation [89,90]. It is important to note that vaccination recommendations differ between countries, so this review draws on recommendations for the depicted pathogens from the German authorities and societies (Table 1) [91–94].





Pathogen	Type of vaccine	Vaccination schedule	Special considerations
Influenza	Inactivated virus	Annually before infection season	Elevated immunogenicity following HD vaccine
SARS-CoV-2	mRNA/vector	Booster depending on new variants	Basic immunization <sup>a</sup> prior to transplantation
Pneumococcus	20-valent conjugate (PCV20)	Depending on vaccination status with PCV13 and PPSV23	No sequential vaccination needed after PCV20
Respiratory syncytial virus	Adjuvanted protein-based, protein-based, mRNA	Single dose as soon as available	No data about immunogenicity nor about timepoint of booster vaccination

 Table 1. Summary of key vaccines against major respiratory pathogens, including vaccine type, target groups, and recommended schedules according to Standing Committee on Vaccination (STIKO)

<sup>a</sup>At least three antigen contacts consisting of at least two vaccines.

HD, high-dose; mRNA, messenger RNA.

Results from a Dutch cohort revealed partial humoral immune response in lung transplanted patients after revaccination with polysaccharide vaccine PPSV23 [95]. In contrast, sequential vaccination with a 23-valent polysaccharide vaccines after a 13-valent conjugate vaccine did not improve antibody response [96]. Humoral immune response after vaccination against Sars-CoV-2 is generally low after transplantation but is increased after repetitive administration of mRNA vaccine in solid organ transplanted patients [97,98]. However, the percentage of patients with sufficient humoral immune response is lower than in immunocompetent individuals [97]. Some immunosuppressants, for example, mycophenolate mofetil in particular has a negative effect on the immune response [97,99]. Despite attenuated humoral immune response cellular-mediated immunity expected protection may co-exist [97].

Since 2023, there are vaccines available against RSV showing promising results to protect older adults 60+ [100,101], recently also a mRNA-based vaccine [102]. Despite pending approval for younger patients, several German specialist societies also recommend widespread use in patients with preexisting pulmonary diseases [103].

# Prophylactic drug therapy against opportunistic pathogens

Lifelong or temporary oral prophylactic therapy has proven to be useful in preventing opportunistic infections, mainly due to cytomegalovirus, PjP and fungal infections.

Introduction of lifelong prophylaxis with trimethoprim–sulfamethoxazole ensured a dramatic drop of severe and potential life-threatening PjP [104]. In case of intolerance or severe side effects atovaquone or inhaled pentamidine can be considered as an alternative therapy [73].

Several options are available for the prophylaxis of invasive aspergillosis, including inhaled amphotericin

B and systemic azoles. Liposomal amphotericin B is better tolerated than conventional amphotericin B formulation and can be applied 3-times weekly but is off-label. Azol therapy is convenient but may have side effects, interactions and is off-label in most countries for prophylaxis after lung transplantation. Due to direct application into the lung allograft, inhalative prophylaxis avoids systemic adverse effects and drugdrug interaction but possibly causes local effects like cough or bronchospasm. Additionally, prophylactic effects are limited to the lung allograft [105,106]. Among systemic azoles, data reveal a sufficient reduction of invasive fungal infection whereas itraconazole and posaconazole have a more favorable safety profile than voriconazole [107–109]. Results mainly based on small sample sizes in retrospective studies underlying the need for further exploration to establish an international standard [105].

The length of CMV prophylaxis depends on the individual risk profile, which is determined by the serostatus of the recipient and the donor. A positive donor (D+) and negative recipient (R-) are considered a high-risk profile with a recommendation for oral prophylaxis with valganciclovir for up to 12 months, whereas this can be shortened to 3–6 months if the recipient (R+) is positive [110–112].

## CONCLUSION

Lung transplant recipients are particularly vulnerable to viral and bacterial infections due to their weakened immune systems. These infections can lead to severe complications, such as pneumonia, graft dysfunction, and increased mortality. Diagnosis is often complex and involves various tests, while treatment can be challenging due to interactions with the immunosuppressive drugs required to prevent organ rejection. Preventive strategies, such as vaccination and the use of prophylactic medications, are essential in reducing infection risks. Despite these measures, infections remain a significant cause of complications and can severely impact the long-term success of lung transplants.

Looking ahead, improving infection management in lung transplant patients will require advancements in diagnostic tools, more effective treatments with fewer side effects, and enhanced vaccination strategies that provide stronger protection. Further research is also needed to develop standardized protocols for preventing and treating infections, ensuring better outcomes and quality of life for transplant recipients.

## Acknowledgements

None.

## **Financial support and sponsorship**

None.

## **Conflicts of interest**

There are no conflicts of interest.

## REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

of special interest

- of outstanding interest
  - Nosotti M, Tarsia P, Morlacchi LC. Infections after lung transplantation. J Thorac Dis 2018; 10:3849–3868.
  - de Zwart A, Riezebos-Brilman A, Lunter G, et al. Respiratory syncytial virus, human metapneumovirus, and parainfluenza virus infections in lung transplant recipients: a systematic review of outcomes and treatment strategies. Clin Infect Dis 2022; 74:2252–2260.
  - Permpalung N, Liang T, Gopinath S, et al. Invasive fungal infections after respiratory viral infections in lung transplant recipients are associated with lung allograft failure and chronic lung allograft dysfunction within 1 year. J Heart Lung Transplant 2023; 42:953–963.
- 4. Dellgren G, Lund TK, Raivio P, et al. Effect of once-per-day tacrolimus
- versus twice-per-day ciclosporin on 3-year incidence of chronic lung allograft dysfunction after lung transplantation in Scandinavia (Scan-CLAD): a multicentre randomised controlled trial. Lancet Respir Med 2024; 12:34–44.

Valuable data about the use of  $\ensuremath{\mathsf{CNI}}$  immunosuppression in lung transplant recipients.

- Marriott DJ, Orla Morrissey C. Common infections following lung transplantation. Esse Lung Transplant 2018: 173–219. doi: 10.1007/978-3-319-90933-2\_15.
- Joean O, Welte T, Gottlieb J. Chest infections after lung transplantation. Chest 2022; 161:937–948.
- Gottlieb J. Community-acquired respiratory viruses. Curr Opin Organ Transplant 2019; 24:311–317.
- Speich R, van der Bij W. Epidemiology and management of infections after lung transplantation. Clin Infect Dis 2001; 33(Suppl 1):S58–S65.
- Garbino J, Soccal PM, Aubert JD, et al. Respiratory viruses in bronchoalveolar lavage: a hospital-based cohort study in adults. Thorax 2009; 64:399–404.
- Gottlieb J, Schulz TF, Welte T, et al. Community-acquired respiratory viral infections in lung transplant recipients: a single season cohort study. Transplantation 2009; 87:1530–1537.
- Kumar D, Husain S, Chen MH, et al. A prospective molecular surveillance study evaluating the clinical impact of community-acquired respiratory viruses in lung transplant recipients. Transplantation 2010; 89:1028–1033.
- de Zwart AES, Riezebos-Brilman A, Alffenaar JC, et al. Evaluation of 10 years of parainfluenza virus, human metapneumovirus, and respiratory syncytial virus infections in lung transplant recipients. Am J Transplant 2020; 20: 3529–3537.
- Chakinala MM, Walter MJ. Community acquired respiratory viral infections after lung transplantation: clinical features and long-term consequences. Semin Thorac Cardiovasc Surg 2004; 16:342–349.

- Peghin M, Los-Arcos I, Hirsch HH, et al. Community-acquired respiratory viruses are a risk factor for chronic lung allograft dysfunction. Clin Infect Dis 2019; 69:1192–1197.
- Allyn PR, Duffy EL, Humphries RM, et al. Graft loss and CLAD-onset is hastened by viral pneumonia after lung transplantation. Transplantation 2016; 100:2424–2431.
- Sayah DM, Koff JL, Leard LE, et al. Rhinovirus and other respiratory viruses exert different effects on lung allograft function that are not mediated through acute rejection. Clin Transplant 2013; 27:E64–71.
- Vandervest KM, Zamora MR. Respiratory viral infections postlung transplantation. Curr Respir Care Rep 2012; 1:162–167.
- Ko JP, Shepard JA, Sproule MW, et al. CT manifestations of respiratory syncytial virus infection in lung transplant recipients. J Comput Assist Tomogr 2000; 24:235–241.
- Matar LD, McAdams HP, Palmer SM, et al. Respiratory viral infections in lung transplant recipients: radiologic findings with clinical correlation. Radiology 1999; 213:735–742.
- Manuel O, Estabrook M. RNA respiratory viral infections in solid organ transplant recipients: guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. Clin Transplant 2019; 33: e13511.
- Rademacher J, Suhling H, Greer M, et al. Safety and efficacy of outpatient bronchoscopy in lung transplant recipients – a single centre analysis of 3197 procedures. Transplant Res 2014; 3:11.
- 22. Azadeh N, Sakata KK, Brighton AM, et al. FilmArray respiratory panel assay: comparison of nasopharyngeal swabs and bronchoalveolar lavage samples. J Clin Microbiol 2015; 53:3784–3787.
- Hammond SP, Gagne LS, Stock SR, et al. Respiratory virus detection in immunocompromised patients with FilmArray respiratory panel compared to conventional methods. J Clin Microbiol 2012; 50:3216–3221.
- Azadeh N, Sakata KK, Saeed A, et al. Comparison of respiratory pathogen detection in upper versus lower respiratory tract samples using the BioFire FilmArray respiratory panel in the immunocompromised host. Can Respir J 2018; 2018:2685723.
- 25. Bouzid D, Hingrat QL, Salipante F, et al. Agreement of respiratory viruses'
   detection between nasopharyngeal swab and bronchoalveolar lavage in
- adults admitted for pneumonia: a retrospective study. Clin Microbiol Infect 2023; 29:942.e1–942.e6..

A retrospective study demonstrating a good agreement between nasopharyngeal swabs and BAL in detecting respiratory viruses among adult patients with suspected pneumonia.

- 26. Kayser MZ, Seeliger B, Valtin C, et al. Clinical decision making is improved by
- BioFire Pneumonia Plus in suspected lower respiratory tract infection after lung transplantation: results of the prospective DBATE-IT study. Transpl Infect Dis 2022; 24:e13725.

Point of care tests offered faster test results compared to conventional tests with good concordance in lung transplant patients with respiratory tract infections.

27. Hinze CA, Lennartz FN, Gras JC, *et al.* A rapid point-of-care polymerase
 chain reaction test in suspected viral respiratory tract infection after lung transplantation – a single-center experience. Transpl Infect Dis 2024; 26: e14349.

Point of care tests in swabs has a good negative predictive value for CARV. BAL is still recommended if swab results are negative and BAL is necessary to assess for bacterial co-infection in immunosuppressed patients.

 28. Yamanaga S, Shimata K, Ohfuji S, *et al.* Excess mortality in COVID-19affected solid organ transplant recipients across the pandemic. Am J Transplant 2024; 24:1495–1508.

This publication provides valuable data about the vulnerability of organ transplanted patients due to COVID-19.

29. Gottlieb J, Simon S, Barton J, *et al.* Efficacy of preexposure prophylaxis to
prevent SARS-CoV-2 infection after lung transplantation: a two center cohort
study during the omicron era. Infection 2023; 51:1481–1489.

Despite being at higher risk for worse outcome severity of COVID-19 and associated mortality were similar in patients with and without preexposure prophylaxis.

- Kamp JC, Hinrichs JB, Fuge J, et al. COVID-19 in lung transplant recipients risk prediction and outcomes. PLoS One 2021; 16:e0257807.
- 31. Solera JT, Árbol BG, Bahinskaya I, et al. Short-course early outpatient remdesivir prevents severe disease due to COVID-19 in organ transplant recipients during the omicron BA.2 wave. Am J Transplant 2023; 23:78–83.
- 32. Solera JT, Árbol BG, Mittal A, *et al.* Longitudinal outcomes of COVID-19 in
   solid organ transplant recipients from 2020 to. Am J Transplant 2024; 24:1303–1316.

The study concludes that COVID-19 severity decreased across different variants in solid organ transplantation. Lung transplantation was associated with worse outcomes and may benefit more from preventive and early therapeutic interventions.

- 33. Kumar D, Michaels MG, Morris MI, et al. Outcomes from pandemic influenza A H1N1 infection in recipients of solid-organ transplants: a multicentre cohort study. Lancet Infect Dis 2010; 10:521–526.
- **34.** Ison MG, Sharma A, Shepard JA, *et al.* Outcome of influenza infection managed with oseltamivir in lung transplant recipients. J Heart Lung Transplant 2008; 27:282–288.

- Kumar D, Ferreira VH, Blumberg E, et al. A 5-year prospective multicenter evaluation of influenza infection in transplant recipients. Clin Infect Dis 2018; 67:1322–1329.
- Edelstein GE, Boucau J, Uddin R, et al. SARS-CoV-2 virologic rebound with nirmatrelvir-ritonavir therapy: an observational study. Ann Intern Med 2023; 176:1577–1585.
- Pelaez A, Lyon GM, Force SD, et al. Efficacy of oral ribavirin in lung transplant patients with respiratory syncytial virus lower respiratory tract infection. J Heart Lung Transplant 2009; 28:67–71.
- Fuehner T, Dierich M, Duesberg C, et al. Single-centre experience with oral ribavirin in lung transplant recipients with paramyxovirus infections. Antivir Ther 2011; 16:733–740.
- 39. Garcia B, Sharma N, Johnson K, et al. Clinical outcomes of paramyxovirus infections in lung transplant recipients treated with oral ribavirin: a two-center case series. Exp Clin Transplant 2019; 17:393–397.
- Burrows FS, Carlos LM, Benzimra M, et al. Oral ribavirin for respiratory syncytial virus infection after lung transplantation: efficacy and cost-efficiency. J Heart Lung Transplant 2015; 34:958–962.
   Gottlieb J, Torres F, Haddad T, et al. A randomized controlled trial of
- Gottlieb J, Torres F, Haddad T, *et al.* A randomized controlled trial of presatovir for respiratory syncytial virus after lung transplant. J Heart Lung Transplant 2023; 42:908–916.
- 42. Gottlieb J, Zamora MR, Hodges T, et al. ALN-RSV01 for prevention of bronchiolitis obliterans syndrome after respiratory syncytial virus infection in lung transplant recipients. J Heart Lung Transplant 2016; 35:213–221.
- 43. Kotton CN, Torre-Cisneros J, Aguado JM, et al. Cytomegalovirus in the transplant setting: Where are we now and what happens next? A report from the International CMV Symposium 2021. Transpl Infect Dis 2022; 24: e13977.
- Balthesen M, Messerle M, Reddehase MJ. Lungs are a major organ site of cytomegalovirus latency and recurrence. J Virol 1993; 67:5360–5366.
- 45. Paraskeva M, Bailey M, Levvey BJ, *et al.* Cytomegalovirus replication within the lung allograft is associated with bronchiolitis obliterans syndrome. Am J Transplant 2011; 11:2190–2196.
- 46. Johansson I, Mårtensson G, Nyström U, et al. Lower incidence of CMV infection and acute rejections with valganciclovir prophylaxis in lung transplant recipients. BMC Infect Dis 2013; 13:582.
- 47. Solans EP, Yong S, Husain AN, et al. Bronchioloalveolar lavage in the diagnosis of CMV pneumonitis in lung transplant recipients: an immunocytochemical study. Diagn Cytopathol 1997; 16:350–352.
- Lodding IP, Schultz HH, Jensen JU, *et al.* Cytomegalovirus viral load in bronchoalveolar lavage to diagnose lung transplant associated CMV pneumonia. Transplantation 2018; 102:326–332.
- 49. Kotton CN, Kumar D, Caliendo AM, et al. The third international consensus guidelines on the management of cytomegalovirus in solid-organ transplantation. Transplantation 2018; 102:900–931.
- Aguilar-Guisado M, Givaldá J, Ussetti P, et al. Pneumonia after lung transplantation in the RESITRA cohort: a multicenter prospective study. Am J Transplant 2007; 7:1989–1996.
- Alsaeed M, Husain S. Infections in heart and lung transplant recipients. Crit Care Clin 2019; 35:75–93.
- He X, Dai HP, Chen QR, et al. Pneumonia relevant to lung transplantation and pathogen distribution. Chin Med J (Engl) 2013; 126:3209–3214.
- 53. Tejada S, Campogiani L, Mazo C, et al. Acute respiratory failure among lung transplant adults requiring intensive care: changing spectrum of causative organisms and impact of procalcitonin test in the diagnostic workup. Transpl Infect Dis 2020; 22:e13346.
- Wojarski J, Ochman M, Medrala W, *et al.* Bacterial infections during hospital stay and their impact on mortality after lung transplantation: a single-center study. Transplant Proc 2018; 50:2064–2069.
- 55. Rezahosseini O, Møller DL, Sørensen SS, et al. An observational prospective cohort study of incidence and outcome of *Streptococcus pneumoniae* and *Hemophilus influenzae* infections in adult solid organ transplant Recipients. Microorganisms 2021; 9:1371.
- Walti LN, Mugglin C, Mombelli M, et al. Vaccine-preventable infections among solid organ transplant recipients in Switzerland. JAMA Netw Open 2023; 6:e2310687.
- de Bruyn G, Whelan TP, Mulligan MS, et al. Invasive pneumococcal infections in adult lung transplant recipients. Am J Transplant 2004; 4:1366– 1371.
- Omori K, Kitagawa H, Nagaoka R, et al. Lung and cerebral nocardiosis caused by *Nocardia elegans* in a lung transplant recipient: a case report and literature review. Intern Med 2023; 62:431–437.
- **59.** Peleg AY, Husain S, Qureshi ZA, *et al.* Risk factors, clinical characteristics, and outcome of Nocardia infection in organ transplant recipients: a matched case-control study. Clin Infect Dis 2007; 44:1307–1314.
- Hemmersbach-Miller M, Stout JE, Woodworth MH, et al. Nocardia infections in the transplanted host. Transpl Infect Dis 2018; 20:e12902.
- **61.** Yetmar ZA, Challener DW, Seville MT, *et al.* Outcomes of nocardiosis and treatment of disseminated infection in solid organ transplant recipients. Transplantation 2023; 107:782–791.
- Coussement J, Lebeaux D, Rouzaud C, Lortholary O. Nocardia infections in solid organ and hematopoietic stem cell transplant recipients. Curr Opin Infect Dis 2017; 30:545–551.

- 63. Restrepo A, Clark NM. Nocardia infections in solid organ transplantation: guidelines from the Infectious Diseases Community of Practice of the American Society of Transplantation. Clin Transplant 2019; 33:e13509.
- Friedman DZP, Doucette K. Mycobacteria: selection of transplant candidates and postlung transplant outcomes. Semin Respir Crit Care Med 2021; 42:460–470.
- 65. Tissot A, Thomas MF, Corris PA, Brodlie M. NonTuberculous Mycobacteria infection and lung transplantation in cystic fibrosis: a worldwide survey of clinical practice. BMC Pulm Med 2018; 18:86.
- 66. Griffith DE, Aksamit T, Brown-Elliott BA, et al. An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. Am J Respir Crit Care Med 2007; 175:367–416.
- Shah SK, McAnally KJ, Seoane L, et al. Analysis of pulmonary nontuberculous mycobacterial infections after lung transplantation. Transpl Infect Dis 2016; 18:585–591.
- Knoll BM, Kappagoda S, Gill RR, et al. Nontuberculous mycobacterial infection among lung transplant recipients: a 15-year cohort study. Transpl Infect Dis 2012; 14:452–460.
- 69. Daley CL, laccarino JM, Lange C, et al. Treatment of nontuberculous mycobacterial pulmonary disease: an official ATS/ERS/ESCMID/IDSA Clinical Practice Guideline. Clin Infect Dis 2020; 71:e1–e36.
- Schmidt JJ, Lueck C, Ziesing S, et al. Clinical course, treatment and outcome of Pneumocystis pneumonia in immunocompromised adults: a retrospective analysis over 17 years. Crit Care 2018; 22:307.
- Fishman JA. Prevention of infection caused by *Pneumocystis carinii* in transplant recipients. Clin Infect Dis 2001; 33:1397–1405.
- Delbove A, Alami H, Tissot A, et al. Pneumocystis pneumonia after lung transplantation: a retrospective multicenter study. Respir Med 2020; 169: 106019.
- Martin SI, Fishman JA. Pneumocystis pneumonia in solid organ transplantation. Am J Transplant 2013; 13(Suppl 4):272–279.
- 74. Iriart X, Challan Belval T, Fillaux J, et al. Risk factors of Pneumocystis pneumonia in solid organ recipients in the era of the common use of posttransplantation prophylaxis. Am J Transplant 2015; 15:190–199.
- 75. Kanne JP, Yandow DR, Meyer CA. Pneumocystis jiroveci pneumonia: highresolution CT findings in patients with and without HIV infection. AJR Am J Roentgenol 2012; 198:W555–W561.
- Bateman M, Oladele R, Kolls JK. Diagnosing *Pneumocystis jirovecii* pneumonia: a review of current methods and novel approaches. Med Mycol 2020; 58:1015–1028.
- Karageorgopoulos DE, Qu JM, Korbila IP, *et al.* Accuracy of β-D-glucan for the diagnosis of *Pneumocystis jirovecii* pneumonia: a meta-analysis. Clin Microbiol Infect 2013; 19:39–49.
- 78. Yasuoka A, Tachikawa N, Shimada K, et al. (1->3) beta-D-glucan as a quantitative serological marker for *Pneumocystis carinii* pneumonia. Clin Diagn Lab Immunol 1996; 3:197–199.
- 79. Asai N, Motojima S, Ohkuni Y, et al. Early diagnosis and treatment are crucial for the survival of *Pneumocystis pneumonia* patients without human immunodeficiency virus infection. J Infect Chemother 2012; 18:898–905.
- 80. Segal BH. Aspergillosis. N Engl J Med 2009; 360:1870-1884.
- Steinbach WJ, Marr KA, Anaissie EJ, et al. Clinical epidemiology of 960 patients with invasive aspergillosis from the PATH Alliance registry. J Infect 2012; 65:453–464.
- 82. Pappas PG, Alexander BD, Andes DR, et al. Invasive fungal infections among organ transplant recipients: results of the Transplant-Associated Infection Surveillance Network (TRANSNET). Clin Infect Dis 2010; 50: 1101–1111.
- De Mol W, Bos S, Beeckmans H, et al. Antifungal prophylaxis after lung transplantation: where are we now? Transplantation 2021; 105:2538–2545.
- 84. Singh N, Husain S. Aspergillus infections after lung transplantation: clinical differences in type of transplant and implications for management. J Heart Lung Transplant 2003; 22:258–266.
- 85. Greene RE, Schlamm HT, Oestmann JW, et al. Imaging findings in acute invasive pulmonary aspergillosis: clinical significance of the halo sign. Clin Infect Dis 2007; 44:373–379.
- Russo A, Tiseo G, Falcone M, Menichetti F. Pulmonary aspergillosis: an evolving challenge for diagnosis and treatment. Infect Dis Ther 2020; 9:511–524.
- Patterson TF, Thompson GR 3rd, *et al.* Practice guidelines for the diagnosis and management of aspergillosis: 2016 update by the Infectious Diseases Society of America. Clin Infect Dis 2016; 63:e1–e60.
- Hamandi B, Fegbeutel C, Silveira FP, et al. Voriconazole and squamous cell carcinoma after lung transplantation: a multicenter study. Am J Transplant 2018; 18:113–124.
- Harboe ZB, Hald A, Ekenberg C, et al. Implementation of a vaccination clinic for adult solid organ transplant candidates: a single-center experience. Vaccine 2023; 41:6637–6644.
- **90.** Felzer JR, Finney Rutten LJ, Wi CI, *et al.* Disparities in vaccination rates in solid organ transplant patients. Transpl Infect Dis 2023; 25:e14010.
- 91. Michaelis KSS, Buda S, Garbe E, *et al.* Beschluss und Wissenschaftliche Begründung der Ständigen Impfkommission (STIKO) für die Aktualisierung der Influenza-Impfempfehlung für Personen im Alter von ≥60 Jahren. Epid Bull 2021; 1:3–25.

0951-7375 Copyright © 2025 Wolters Kluwer Health, Inc. All rights reserved.

- 92. Piechotta V KJ, Berner R, Bogdan C, et al. Aktualisierung der COVID-19-Impfempfehlung in den allgemeinen Empfehlungen der STIKO 2024 und die dazugehörige wissenschaftliche Begründung. Epid Bull 2024; 2:3–19.
- 93. Schlaberg F V-BS, Falman A, Wilhelm J, et al. Aktualisierung der Empfehlungen der STIKO zur Standardimpfung von Personen ≥60 Jahre sowie zur Indikationsimpfung von Risikogruppen gegen Pneumokokken und die dazugehörige wissenschaftliche Begründung. Epid Bull 2023; 39:3–44.
- 94. Falman A, Flasche SV, Günther S, et al. Beschluss und Wissenschaftliche Begründung zur Empfehlung der STIKO für eine Standardimpfung gegen Erkrankungen durch Respiratorische Synzytial-Viren (RSV) für Personen ≥ 75 Jahre sowie zur Indikationsimpfung von Personen im Alter von 60 bis 74 Jahren mit Risikofaktoren. Epid Bull 2024; 32:3–28.
- van Kessel DA, Hoffman TW, Kwakkel-van Erp JM, et al. Long-term follow-up of humoral immune status in adult lung transplant recipients. Transplantation 2017; 101:2477–2483.
- 96. Hoffman TW, Meek B, Rijkers GT, et al. Pneumococcal conjugate vaccination followed by pneumococcal polysaccharide vaccination in lung transplant candidates and recipients. Transplant Direct 2020; 6:e555.
- Dauriat G, Beaumont L, Luong Nguyen LB, et al. Efficacy of three COVID-19 vaccine doses in lung transplant recipients: a multicentre cohort study. Eur Respir J 2023; 61:.
- Bárczi E, Varga V, Nagy A, *et al.* Serological findings following the second and third SARS-CoV-2 vaccines in lung transplant recipients. Immun Inflamm Dis 2022; 10:e646.
- Lucca F, Bezzerri V, Danese E, et al. Immunogenicity and safety of the BNT162b2 COVID-19 vaccine in patients with cystic fibrosis with or without lung transplantation. Int J Mol Sci 2023; 24:908.
- 100. Papi A, Ison MG, Langley JM, et al. Respiratory syncytial virus prefusion F protein vaccine in older adults. N Engl J Med 2023; 388:595–608.
- 101. Walsh EE, Pérez Marc G, Zareba AM, et al. Efficacy and safety of a bivalent RSV prefusion f vaccine in older adults. N Engl J Med 2023; 388:1465–1477.
- **102.** Mullard A. FDA approves mRNA-based RSV vaccine. Nat Rev Drug Discov 2024; 23:487.

- 103. Addo M, Cornely O, Denkinger M, et al. RSV vaccination strategies for highrisk patients 2023: a collaborative position paper by leading German medical societies and organizations. Infection 2024; 52:285–288.
- 104. Trubin PA, Azar MM. Current concepts in the diagnosis and management of pneumocystis pneumonia in solid organ transplantation. Infect Dis Clin North Am 2023; 37:617–640.
- 105. Husain S, Camargo JF. Invasive Aspergillosis in solid-organ transplant recipients: guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. Clin Transplant 2019; 33: e13544.
- 106. Drew RH, Dodds Ashley E, Benjamin DK Jr, et al. Comparative safety of amphotericin B lipid complex and amphotericin B deoxycholate as aerosolized antifungal prophylaxis in lung-transplant recipients. Transplantation 2004; 77:232–237.
- 107. Cadena J, Levine DJ, Angel LF, et al. Antifungal prophylaxis with voriconazole or itraconazole in lung transplant recipients: hepatotoxicity and effectiveness. Am J Transplant 2009; 9:2085–2091.
- 108. Jeong W, Snell GI, Levvey BJ, et al. Clinical effectiveness of early posaconazole suspension preemptive therapy in lung transplant recipients: the Alfred's experience. J Antimicrob Chemother 2017; 72:2089–2092.
- **109.** Neoh CF, Snell GI, Levvey B, *et al.* Preemptive treatment with voriconazole in lung transplant recipients. Transpl Infect Dis 2013; 15:344–353.
- 110. Jaksch P, Zweytick B, Kerschner H, et al. Cytomegalovirus prevention in highrisk lung transplant recipients: comparison of 3- vs 12-month valganciclovir therapy. J Heart Lung Transplant 2009; 28:670–675.
- 111. Ruiz-Arabi E, Torre-Ĉisneros J, Aguilera V, et al. Management of cytomegalovirus in adult solid organ transplant patients: GESITRA-IC-SEIMC, CIBER-INFEC, and SET recommendations update. Transplant Rev (Orlando) 2024; 38:100875.
- 112. Monforte V, Sintes H, López-Gallo C, et al. Risk factors, survival, and impact of prophylaxis length in cytomegalovirus-seropositive lung transplant recipients: a prospective, observational, multicenter study. Transpl Infect Dis 2017; 19:. doi: 10.1111/tid.12694.