Contents lists available at ScienceDirect

Maturitas

journal homepage: www.elsevier.com/locate/maturitas

Practice guidelines

Practice guideline on ovarian tissue cryopreservation and transplantation in the prevention and treatment of iatrogenic premature ovarian insufficiency *

ARTICLE INFO

Keywords Ovarian tissue cryopreservation Ovarian tissue transplantation latrogenic premature ovarian insufficiency Prevention and treatment Practice guideline Ovarian function and fertility preservation

ABSTRACT

Premature ovarian insufficiency (POI) refers to the decline of ovarian function before the age of 40. POI causes a reduction in or loss of female fertility, accompanied by different degrees of menopausal symptoms, which increases the risk of chronic diseases related to early menopause and seriously affects patients' quality of life and health. It is conservatively estimated that at least one million prepubertal girls and women of reproductive age in China are at risk of iatrogenic POI caused by radiotherapy and chemotherapy every year. With the development of medical technology and the breakthrough of scientific and technological advances, preventing and treating iatrogenic POI have become possible. International and national guidelines consider cryopreserved ovarian tissue transplantation to be the most promising method of preserving the ovarian function and fertility of prepubertal girls and women of reproductive age who cannot delay radiotherapy and chemotherapy. In order to guide the clinical application of ovarian tissue cryopreservation and transplantation technology in China, the Guideline Working Group finally included 14 scientific questions and 18 recommendations through a questionnaire survey, field investigation, and consultation of a large number of Chinese and English literature databases in order to provide a reference for colleagues in clinical practice.

1. Introduction

Premature ovarian insufficiency (POI) refers to the decline in ovarian function before the age of 40. Traditionally, the prevalence rate of POI is about 1 %. Recently, it has been reported that the prevalence rate of POI has increased significantly, reaching $3.7 \% \sim 10 \%$ [1,2]. POI can cause the decrease or loss of female fertility, accompanied by different degrees of peri-menopausal symptoms. In addition, the fluctuation of estrogen levels can increase the risk of cardiovascular diseases, osteoporosis, Alzheimer's disease and other diseases, significantly shorten life expectancy and seriously affect patients' quality of life and health [3]. Iatrogenic POI refers to ovarian dysfunction related to medical care behaviour. The common causes are surgery, chemotherapy, radiotherapy and intake of other ovarian toxic substances, accounting for about 50 % of POI [4,5].

According to data from the World Health Organization (WHO), there are >4.5 million new cancers in China annually. It is conservatively estimated that at least one million prepubertal girls and women of reproductive age in China are at risk of iatrogenic POI due to radiotherapy and chemotherapy every year and should be evaluated to preserve ovarian function and fertility. Therefore, it is very important to prevent the occurrence of iatrogenic POI effectively [6]. It is internationally agreed that ovarian tissue cryopreservation and transplantation (OTCT) is the only method to preserve ovarian function and fertility in children and women of reproductive age who cannot delay radiotherapy and chemotherapy, and it is also the most effective and promising method to preserve ovarian function and fertility at present [7,8]. Multidisciplinary experts were asked to join a working group to formulate this guideline to better guide the clinical application of OTCT in the prevention and treatment of iatrogenic POI.

The guideline working group collected clinical issues of concern to clinicians and patients through questionnaires. The survey collected 500 doctors' and 300 patients' questionnaires from 34 provinces, autonomous regions and municipalities directly under the Central Government and combined them to remove duplicates. According to their importance ranking and the feasibility of literature retrieval, the guideline working group finally decided to include 14 clinical questions after further discussion. This guideline comprises 14 clinical questions and 18 recommendations. The evidence level (Table 1) and recommendation level (Table 2) refer to the standards of the Oxford Center for Evidence-based Medicine in 2009 [9] and are marked in brackets after each recommendation.

https://doi.org/10.1016/j.maturitas.2024.107922

Received 28 September 2023; Received in revised form 17 January 2024; Accepted 19 January 2024 Available online 26 January 2024







^{*} Xiangyan Ruan and Committee of Fertility Preservation and Preservation of China Association for the Promotion of Health Science and Technology, Chinese Society of Gynecological Endocrinology affiliated to the International Society of Gynecological Endocrinology, Society Endocrinology Branch of Beijing Institute of Obstetrics & Gynecology, Society on Fertility Preservation affiliated to Chinese Preventive Medicine Association, Reproductive Endocrine Professional Committee of China Maternal and Child Health Research Association.

^{0378-5122/© 2024} The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC license (http://creativecommons.org/licenses/by-nc/4.0/).

Table 1

The level and meaning of evidence used in this guide.

Level of evidence	Meaning
1a	Homogeneous randomized controlled trials, initial cohort, prospective cohort, systematic review of level 1 diagnostic studies, multi-population based studies, multicenter studies
1b	Randomized controlled trial with a narrow confidence interval, follow-up $>$ 80 %, single population, single-center study
1c	Evidence that the observation result is "all or no."
2a	A systematic review of homogeneous cohort studies, retrospective cohorts of randomized controlled studies, and diagnostic studies above grade 2
2b	1 cohort (including low-quality randomized controlled studies), retrospective cohort, poor follow-up
2c	Outcome study
3a	Systematic review of homogeneous case-control studies
3b	1 case-control study, discontinuous study
4	Case series study, low-quality cohort study, or case-control study
5	Expert opinion, no clear, rigorous evaluation, basic research based on laboratory

Table 2

Recommended levels and meanings used in this guide.

Recommended grade	Meaning
А	Consistent Level 1 evidence
В	Consistent Level 2 or Level 3 evidence or inference of Level 1 evidence
С	Level 4 evidence, or level 2 or level 3 evidence inference
D	Level 5 evidence, or inconsistent or uncertain conclusions

2. Ovarian tissue cryopreservation

2.1. Which population is suitable for ovarian tissue cryopreservation?

Ovarian tissue cryopreservation (OTC) refers to removal of ovarian tissue followed by cryopreservation based on the principle of cryobiology. It is the only choice to preserve ovarian function and fertility in prepubertal girls and women of reproductive age who cannot delay radiotherapy and chemotherapy. It does not require ovarian stimulation and delay follow-up treatment, preserves thousands of follicles at one time, preserves fertility, and restores ovarian endocrine function to a certain extent [10,11]. OTCT has been formally used in clinical practice and is no longer an experimental technology, which is the main method to prevent and treat iatrogenic POI [7]. If the patient has received low toxicity and low-dose chemotherapy, the ovarian function of the patient should be evaluated to judge whether OTC can be performed, and chemotherapy before OTC should not be regarded as a contraindication [7,12,13].

Recommendation 1–1: Ovarian tissue cryopreservation is suitable for preserving ovarian function and fertility in patients with tumour and non-tumour diseases and is the only preservation method of ovarian function and fertility for prepubertal girls and women of reproductive age who cannot delay ovarian toxicity treatment. (2b, B).

Recommendation 1–2: Chemotherapy before OTC should not be considered a contraindication to ovarian tissue cryopreservation. (2b, B).

2.2. What are the cautions for ovarian tissue sampling surgery?

In principle, an ovarian tissue biopsy should be taken before radiotherapy and chemotherapy or at the same time as pelvic surgery. If the patient has received low toxicity and low-dose chemotherapy, the ovarian function of the patient should be evaluated to judge whether OTC can be performed. The sampling plan is made individually according to the risk of iatrogenic POI and ovarian reserve function. Generally, at least 1/2 or 2/3 of one ovary or 1/2 of both ovaries are biopsied. Because of the small size of the ovary in prepubertal girls, a unilateral oophorectomy is recommended.

It is suggested that qualified units should use a laparoscope to biopsy ovarian tissue or perform biopsy at the same time as laparotomy, the corpus luteum should be avoided as much as possible, a cold knife should be used, surgical energy devices should not come into contact with the ovary, thermal injury should be avoided and the integrity of the biopsied ovarian tissue should be ensured. Surgical energy devices can be used in children when removing a unilateral ovary, but heat injury must be eliminated and ischemic and mechanical injuries such as clamping and extrusion for a long time should be avoided.

To avoid warm ischemia injury, ovarian tissue should be put into hypothermic transfer fluid immediately. The hospitalization time of laparoscopic ovarian tissue sampling surgery ranges from 1 to 3 days. There is no report of intraoperative and postoperative ovarian tissue sampling surgery complications, which can be managed according to general laparoscopic examination [10,14-17].

Recommendation 1–3: Ischemia, mechanical injury and thermal injury must be avoided during ovarian tissue biopsy. (1c, A).

2.3. Which freezing method should be used for ovarian tissue cryopreservation?

OTC methods include slow freezing and vitrification. At present, the standard method of OTC worldwide is slow freezing [7,10,12,18]. From over 200 babies born after OTCT worldwide, only two live births are realized by vitrification of ovarian tissue [18,19].

Recommendation 1–4: Slow-freezing is the standard method of ovarian tissue cryopreservation, and vitrification of ovarian tissue is still a clinical study. (1a, A).

3. Ovarian tissue transplantation

3.1. Is there a risk of recurrence of primary disease in patients with malignant tumours undergoing ovarian tissue transplantation?

Gellert et al. reported monitoring ovarian tissue transplantation from 21 countries; no disease recurrence was caused by ovarian tissue transplantation [20]. According to the data from five European centers in 2021, there is no correlation between the recurrence risk of primary disease and OTCT [12]. For patients with a high risk of ovarian contamination such as leukemia, OTC after chemotherapy to complete remission can significantly reduce the risk of ovarian contamination [21–24].

Evaluating the safety of patients with malignant tumours before ovarian tissue transplantation is very important. Histological microscopy, immunohistochemical staining, fluorescence in situ hybridization (FISH), real-time fluorescence quantitative reverse transcription polymerase chain reaction, etc., can screen specific markers of primary tumours at the cellular or molecular level. For patients lacking specific tumour markers, multicolour flow cytometry can also be selected to detect ovarian tissue. Xenotransplantation experiments in immunodeficient mice are important methods for evaluating the recurrence risk of ovarian tissue implantation in tumour patients, which can evaluate the potential risk of tumour recurrence. The above assessment methods provide risk assessment for tumour patients with a high risk of ovarian contamination [25–27]. After ovarian tissue transplantation, besides the follow-up of ovarian function recovery, the primary disease should be followed up routinely.

In ovarian tissue preparation, immature oocytes in ovarian tissue are cultured in-vitro and cryopreserved. This can improve the fertility preservation efficiency of patients and is also a safe choice for patients with a high risk of ovarian contamination [28–31].

Recommendation 2–1: The risk of recurrence of primary disease after cryopreserved ovarian tissue transplantation is very low. (3b, B).

Recommendation 2–2: Safety assessment of tumour contamination should be carried out before cryopreserved ovarian tissue transplantation. (3b, B).

Recommendation 2–3: For patients with a high risk of cryopreserved ovarian tissue transplantation such as leukemia, cryopreservation of ovarian tissue after chemotherapy to complete remission can reduce the risk of cryopreserved ovarian tissue transplantation. (4, C).

3.2. How should the timing and indication of ovarian tissue transplantation be determined?

Transplantation time should be determined according to whether the patient's primary disease is cured, whether clinical rehabilitation is performed, whether full communication with the patient is possible, and what the individual situation is after multi-disciplinary consultation. Generally, the primary disease is relieved, and patients have perimenopausal symptoms related to ovarian function decline, such as hot flushes and sweating, serum follicle-stimulating hormone (FSH) ≥ 25 IU/L or serum anti-Muellerian hormone (AMH) < 1.1 ng/mL. After stop of ovarian toxicity treatment for at least 3–6 months, ovarian tissue transplantation can be considered [10,11].

Recommendation 2–4: Ovarian tissue transplantation should be considered at least 3–6 months after completion of radiotherapy and chemotherapy for tumour patients. (2b, B).

Recommendation 2–5: The timing of ovarian tissue transplantation should be decided after a multi-disciplinary consultation. (4, C).

3.3. Can transplantation of cryopreserved ovarian tissue restore ovarian function?

Gellert et al. reported that ovarian function recovered in 95 % of the 318 OTCT women worldwide [20]. 285 women from five European centers were followed up after OTCT, and the recovery rate of ovarian endocrine function in the center with the highest recovery rate was 97 % [12]. OTCT has been performed in 19 patients in China's first ovarian tissue cryobank, and the recovery rate of ovarian function after surgery is as high as 100 % [32].

A meta-analysis showed that the median time for ovarian function to return to normal was 19 weeks [19]. In China's first ovarian tissue cryobank, all patients who underwent OTCT recovered their ovarian function within 3–4 months [32].

Before and after cryopreserved ovarian tissue transplantation, ovarian function in patients with peri-menopausal and menopausalrelated symptoms due to POI may not have returned to normal. If there is no contraindication, menopausal hormone therapy (MHT) can be considered. Traditional Chinese medicine is widely used in clinics as one of the drug therapies for POI. Those with MHT contraindications or fear of MHT can reasonably choose traditional Chinese medicine, such as Kuntai capsules. Alternative therapies, such as botanical drugs, can also be selected according to the patient's situation [32,33].

Recommendation 2–6: Cryopreserved ovarian tissue transplantation can restore ovarian function. (2b, B).

3.4. Can cryopreserved ovarian tissue transplantation be used to induce puberty development?

Impaired ovarian function and decreased fertility are among the most important long-term complications of prepubertal cancer treatment [34]. According to statistics, the 5-year survival rate of childhood cancer can reach >85 %, and OTC is the only fertility preservation method for children [6]. Poirot et al. reported the world's first case of retransplanting cryopreserved ovarian tissue back into the body to induce puberty. Patients underwent cryopreserved ovarian tissue transplantation 27 months after hematopoietic stem cell transplantation (HSCT). Secondary sexual characteristics began to appear two months after transplantation and menarche appeared eight months after

transplantation [35]. Since then, there have been many related reports of different diseases [36–38]. Current data show that the recovery rate, pregnancy rate and live birth rate of ovarian tissue cryopreservation before 18 years old are not lower than those undergoing cryopreservation after 18 years of age [39]. It is feasible for prepubertal girls to induce puberty and obtain fertility by OTCT. Data on the duration of induced puberty following OTCT are limited. There have been reports of transplantation of cryopreserved ovarian tissue in childhood and live births after pregnancy [37,40,41].

Recommendation 2–7: Cryopreserved ovarian tissue transplantation can be used to induce puberty development. (4, C).

3.5. Can pregnancy be obtained after transplantation of cryopreserved ovarian tissue?

There are differences in the pregnancy rates of OTCT in different ovarian tissue cryopreservation centers. A meta-analysis of 568 cases of post-OTCT fertility assessment showed a 37 % pregnancy rate after cryopreserved ovarian tissue transplantation [19]. However, the pregnancy rate of 53 cohorts in Denmark after OTCT was 56 % [42]. In 2021, China reported the first successful natural pregnancy and live birth after OTCT [43,44]. The FertiPROTEKT network reported on 244 transplants in 196 patients where 61.7 % of pregnant cases were natural pregnancies, and the pregnancy rate after transplantation decreased with the age of patients when ovarian tissue was cryopreserved [45].

Recommendation 2–8: Patients under 35 with good ovarian reserve have high pregnancy rates after OTCT. (2b, B).

4. Application of cryopreserved ovarian tissue transplantation

4.1. Can some patients with pelvic malignant tumours preserve ovarian function and fertility through ovarian tissue cryopreservation and transplantation?

Patients with early cervical squamous cell carcinoma have a low risk of ovarian metastasis, so patients with cervical cancer are less likely to recur after OTCT. According to the data from the first ovarian tissue cryobank in China, cervical cancer accounts for 49.31 % of OTC cases [46]; half the cases undergoing OTCT are patients with cervical cancer [32]. Cervical cancer accounts for about 61 % of gynecological tumour patients receiving OTC worldwide [20]. OTCT can help cervical cancer patients restore ovarian endocrine function, but successful pregnancy depends on whether the uterus is preserved and the degree of uterus damage caused by radiotherapy and chemotherapy [32].

OTCT can also be considered for patients with early and low-risk endometrial cancer. There have been cases of OTCT for patients with endometrial cancer, which have successfully restored ovarian endocrine function, and there are no reports of tumour recurrence [32].

When patients with ovarian tumours choose OTCT, it is necessary to evaluate the type, grade and stage of tumours and exclude the possibility of ovarian tissue carrying cancer. It is generally believed that ovarian cancer is a contraindication to OTCT. However, in 2015, Dittrich et al. reported a pregnancy and live birth case after OTCT in a patient with stage IIIc G1 ovarian serous adenocarcinoma and removed the transplanted ovarian tissue six weeks after delivery [47].

For other pelvic malignant tumours, such as rectal cancer and bladder cancer patients, the pelvic radiotherapy dose for rectal cancer patients is usually at least 45 Gy, and the pelvic 2 Gy radiotherapy dose can cause 50 % follicle death. Therefore, ovarian tissue should be biopsied and cryopreserved before radiotherapy to preserve ovarian function and fertility [48]. The first ovarian tissue cryobank in China reported on a patient with rectal cancer whose ovarian endocrine function is still good four years after transplantation of cryopreserved ovarian tissue [32].

Recommended 3–1: OTCT be used to restore ovarian function and fertility in patients with pelvic malignant tumours. (4, C).

4.2. Can patients with endometriosis preserve ovarian function and fertility through ovarian tissue cryopreservation and transplantation?

Patients with severe and recurrent ovarian endometriosis have been proven to have impaired ovarian reserve after surgery [49]. At the same time as endometriosis surgery, the healthy ovarian tissue cortex that has not been affected is cryopreserved and can be transplanted back into the body after the patient's disease is cured if the ovarian function is damaged. This provides a more promising choice for the fertility and ovarian function preservation of patients with endometriosis. It has been reported that this technology has been used to restore ovarian endocrine function and pregnancy [50–53].

Recommendation 3–2: Ovarian endometriosis and its surgical treatment will seriously affect ovarian function, and unaffected normal ovarian tissue can be cryopreserved at the time of surgery, which can preserve ovarian function and fertility. (3c, B).

4.3. Can breast cancer patients preserve ovarian function and fertility through ovarian tissue cryopreservation and transplantation?

Breast cancer is one of the best indications of OTCT. Among 2475 OTC patients reported in 2023, breast cancer accounted for 53 % [54]. Histological evaluation of 94 breast cancer patients undergoing OTC showed that follicular activity and density were similar in fresh and frozen-thawed ovarian tissues, and no micrometastases were found in ovarian tissues [55]. After thawing and transplantation of cryopreserved ovarian tissue, the recovery rate of ovarian function is over 95 % [56], and the cumulative live birth rate is about 40 % [57].

The timing of choosing OTCT for breast cancer patients should be determined so that the disease is at a low risk of recurrence and the patient has completed anti-tumour treatment for at least 3–6 months. For breast cancer patients with low recurrence risk who require adjuvant endocrine therapy, after multi-disciplinary evaluation, fully communicating the tumour risk and reproductive needs with patients, and receiving endocrine therapy for at least 2–3 years, they can carefully choose pregnancy after ovarian tissue transplantation. However, it is strongly recommended that they continue to complete endocrine therapy after delivery [58,59].

Recommendation 3–3: Young breast cancer patients can preserve ovarian function and fertility by ovarian tissue cryopreservation and transplantation. (4, C).

4.4. How to preserve the fertility of hematopoietic stem cell transplantation patients?

HSCT is widely used to treat leukemia, lymphoma, and immune diseases with HSCT indications (pigmented villonodular synovitis, dermatomyositis, hyperimmunoglobulin E syndrome, chronic active Epstein-Barr virus infection, etc.), metabolic diseases (mucopolysaccharide storage disease, galactosialic acid storage disease, pyruvate kinase deficiency anemia, mucolipid storage disease, metachromatic leukodystrophy, Niemann-Peak disease, etc.), non-malignant hematological diseases (aplastic anemia, myelodysplastic syndrome, thalassemia, Fanconi anemia, congenital dyserythropoietic anemia, congenital neutropenia, platelet dysfunction). In addition to non-solid tumours, HSCT is also applied to malignant solid tumours such as nephroblastoma and neuroblastoma [60]. However, HSCT can lead to severe iatrogenic POI, and the incidence rate can be as high as 93 % [61,62]. Therefore, all HSCT children and female patients of reproductive age should be provided with a timely fertility preservation consultation. Even after receiving HSCT during childhood, ovarian function and fertility should be evaluated and appropriate intervention should be given to increase the chances of future fertility and reduce the harm of early occurrence of various chronic diseases related to POI [60,63].

Recommendation 3–4: HSCT can lead to iatrogenic POI, and ovarian tissue cryopreservation and transplantation is the first choice to preserve

ovarian function and fertility in such patients. (3c, B).

5. Construction of fertility preservation network

5.1. Why does cryopreservation of ovarian tissue suggest centralization?

OTCT technology is difficult, advanced and innovative. Centers with >10 cases of ovarian tissue transplantation are considered large centers, but there are only over 20 large ovarian tissue cryopreservation centers worldwide [19]. According to international guidelines, ovarian tissue cryopreservation should be centralized to ensure strict and high-standard quality assurance and management, safety and effectiveness. That is, ovarian tissue sampling surgery can be carried out in various places, and the biopsied ovarian tissue can be transported to the cryobank center for cryopreservation and storage [11,64]. For example, the FertiPROTEKT Fertility Preservation Network in Europe [43,65,66] and the American Cancer Fertility Alliance [67,68]. In 2012, Beijing Obstetrics and Gynecology Hospital, Capital Medical University, established China's first ovarian tissue cryobank. Fifty-two medical units have participated in the fertility preservation of China's first clinical ovarian tissue cryobank.

Recommendation 4–1: Ovarian tissue cryopreservation should be centralized. (2b, B).

5.2. Is it feasible to build a wide-area fertility preservation network?

The application of high-tech equipment and standardized process to provide high-quality and high-standard cryopreservation, quality inspection and long-term storage of ovarian tissue for patients all over the country, to ensure a high success rate after thawing and transplantation, and to realize the recovery of ovarian endocrine function has been proved to be effective and feasible [69]. Low-temperature transport method at 4–8 °C is widely used for organ preservation and transport. Low-temperature delays cell metabolism and reduces cell oxygen demand and consumption [70]. In 2018, the European FertiPROTEKT Fertility Preservation Network pointed out that ovarian tissue should be remotely transported to the ovarian tissue cryobank center within 24 h using a 4–8 °C transport box [71].

Recommendation 4–2: Fresh ovarian tissue is transported at 4–8 $^{\circ}$ C for 24 h, which has no effect on follicular activity, and remote transportation is feasible. (4, C).

See the following table for all recommended items in this practice guideline:

Recommended content	Quality of evidence	Recommended strength
1. Ovarian tissue cryopreservation		
Recommendation 1–1: Ovarian tissue cryopreservation is suitable for preserving ovarian function and fertility	2b	В
in patients with tumour and non-tumour diseases and is the only preservation method of ovarian function and fertility for prepubertal girls and women of reproductive age who cannot delay ovarian toxicity treatment.		
Recommendation 1–2: Chemotherapy before OTC should not be considered a contraindication to ovarian tissue cryopreservation.	2b	В
Recommendation 1–3: Ischemia, mechanical injury and thermal injury must be avoided during ovarian tissue biopsy.	1c	Α
Recommendation 1–4: Slow-freezing is the standard method of ovarian tissue cryopreservation, and vitrification of ovarian tissue is still a clinical study. 2. Ovarian tissue transplantation	1a	Α

(continued on next page)

Practice guidelines

(continued)

Recommended content	Quality of evidence	Recommended strength
Recommendation 2–1: The risk of	3b	В
recurrence of primary disease after		
cryopreserved ovarian tissue		
transplantation is very low.		
Recommendation 2–2: Safety assessment	3b	В
of tumour contamination should be		
carried out before cryopreserved ovarian		
tissue transplantation.		
Recommendation 2–3: For patients with a	4	С
high risk of cryopreserved ovarian tissue		
transplantation, such as leukemia,		
cryopreservation of ovarian tissue after		
chemotherapy to complete remission can		
reduce the risk of cryopreserved ovarian		
tissue transplantation.		
Recommendation 2–4: Ovarian tissue	2b	В
transplantation should be considered at	20	2
least 3–6 months after completion of		
radiotherapy and chemotherapy are		
finished for tumour patients.		
Recommendation 2–5: The timing of	4	С
ovarian tissue transplantation should be	•	G
decided after a multi-disciplinary		
consultation.		
Recommendation 2–6: Cryopreserved	2b	В
ovarian tissue transplantation can	20	b
restore ovarian function.		
Recommendation 2–7: Cryopreserved	4	С
ovarian tissue transplantation can be	•	G
used to induce puberty development.		
Recommendation 2–8: Patients under 35	2b	В
years of age with good ovarian reserve	20	2
have high pregnancy rates after OTCT.		
3. The application of ovarian tissue		
cryopreservation transplantation in diseases		
Recommendation 3–1: OTCT be used to	4	С
restore ovarian function and fertility in		
patients with pelvic malignant tumours.		
Recommendation 3–2: Ovarian	3c	В
endometriosis and its surgical treatment		
will seriously affect ovarian function,		
and unaffected normal ovarian tissue		
can be cryopreserved at the time of		
surgery, which can preserve ovarian		
function and fertility.		
Recommendation 3–3: Young breast	4	С
cancer patients can have their ovarian		
function and fertility preserved by		
ovarian tissue cryopreservation and		
transplantation.		
Recommendation 3-4: HSCT can lead to	3c	В
iatrogenic POI, and ovarian tissue		
cryopreservation and transplantation is		
the first choice to preserve ovarian		
function and fertility in such patients.		
4. Construction of a fertility preservation		
network		
Recommendation 4–1: Ovarian tissue	2b	В
cryopreservation should be centralized.		
Recommendation 4-2: Fresh ovarian tissue	4	С
is transported at 4–8 $^\circ\text{C}$ for 24 h, which		
has no effect on follicular activity, and		
remote transportation is feasible.		

6. Discussion

This practice guideline consists of questions and recommendations, with 14 clinical questions and 18 recommendations, written by a multidisciplinary team of gynecologists, fertility specialists, oncologists, reproductive medicine doctors and scientists. For patients, the risks and benefits of fertility preservation interventions require multi-disciplinary discussion and decision-making, and it is essential to provide patients with appropriate information and support their decisions. OTCT is the most effective and promising method to protect ovarian function and fertility. Multi-disciplinary experts have been organized to develop this practice guideline to better guide the clinical application of OTCT technology in preventing and treating iatrogenic POI. Although the guidelines are generally limited, and the evidence supporting the current guidelines is also limited, the expert group believes that the guidelines will contribute to the best practices of OTCT in preventing and treating iatrogenic POI.

Contributors

Xiangyan Ruan contributed to the conception and design of the idea, coordination, and manuscript preparation and editing and was a member of the Expert Panel.

Che Xu prepared the manuscript and was a member of the Expert Panel.

Hefeng Huang was a member of the Expert Panel.

Binghe Xu was a member of the Expert Panel.

Juan Du coordinated the preparation and revision of the manuscript and was a member of the Expert Panel.

Jiaojiao Cheng coordinated the preparation and revision of the manuscript and was a member of the Expert Panel. Fengyu Jin was a member of the Expert Panel. Muqing Gu was a member of the Expert Panel. Weimin Kong was a member of the Expert Panel. Chenghong Yin was a member of the Expert Panel. Yurui Wu was a member of the Expert Panel. Qinjie Tian was a member of the Expert Panel. Yunxia Cao was a member of the Expert Panel. Ruifang Wu was a member of the Expert Panel. Liangzhi Xu was a member of the Expert Panel. Jing Jin was a member of the Expert Panel. Yanglu Li was a member of the Expert Panel. Yinmei Dai was a member of the Expert Panel. Rui Ju was a member of the Expert Panel. Fei Ma was a member of the Expert Panel. Gang Wang was a member of the Expert Panel. Wei Wei was a member of the Expert Panel. Xiaojun Huang was a member of the Expert Panel. Maoquan Qin was a member of the Expert Panel. Yuan Lin was a member of the Expert Panel. Yuan Sun was a member of the Expert Panel. Rong Liu was a member of the Expert Panel. Wei Zhang was a member of the Expert Panel. Xiaodong Li was a member of the Expert Panel. Lin Zou was a member of the Expert Panel. Min Hao was a member of the Expert Panel. Xiyang Ye was a member of the Expert Panel. Fuling Wang was a member of the Expert Panel. Yue Wang was a member of the Expert Panel. Zhuoying Hu was a member of the Expert Panel. Yanhong Huang was a member of the Expert Panel. Tianyuan Zhu was a member of the Expert Panel. Caihong Yang was a member of the Expert Panel. Jinping Wang was a member of the Expert Panel. Xiaomin Yang was a member of the Expert Panel. Rong Ni was a member of the Expert Panel. Liqun Wang was a member of the Expert Panel. Guangxia Luo was a member of the Expert Panel. Aiping Min was a member of the Expert Panel. Siyou Zhang was a member of the Expert Panel. Peiling Li was a member of the Expert Panel. Linghui Cheng was a member of the Expert Panel. Lianfang Li was a member of the Expert Panel. Quanfang Jin was a member of the Expert Panel. Dongmei Shi was a member of the Expert Panel.

Yan Li was a member of the Expert Panel.

Fangying Ren was a member of the Expert Panel.

Yanxiang Cheng was a member of the Expert Panel.

Jumin Niu was a member of the Expert Panel.

Ying Tian was a member of the Expert Panel.

Alfred O. Mueck revised the manuscript and was a member of the Expert Panel.

All authors participated in preparing the practice guidelines and approved the final version of the manuscript.

Funding

This work was supported by the China Association for Promotion of Health Science and Technology Special Fund project for Scientific research (JKHY2020003); Beijing Municipal Health Commission, demonstration construction project of Clinical Research ward (No: BCRW202109); Capital's Funds for Health Improvement and Research (No. 2020-2-2112); Beijing Natural Science Foundation (No. 7202047); Beijing Municipal Administration of Hospitals' Ascent Plan (No. DFL20181401); High-end Foreign Experts Introduction Program Project (G2022001018).

Provenance and peer review

This article was not commissioned and was externally peer reviewed.

Declaration of competing interest

All members of the expert group declare that there is no conflict of interest.

References

- R. Giri, A.J. Vincent, Prevalence and risk factors of premature ovarian insufficiency/early menopause, Semin. Reprod. Med. 38 (4–5) (2020) 237–246 (PMID: 33434933).
- [2] L. Webber, M. Davies, R. Anderson, et al., ESHRE guideline: management of women with premature ovarian insufficiency, Hum. Reprod. 31 (5) (2016) 926–937 (PMID: 27008889).
- [3] R.W. Rebar, C.S. Keator, Expanding our knowledge of premature ovarian insufficiency, Fertil. Steril. 115 (2) (2021) 328–329 (PMID: 33272635).
- [4] M. De Vos, P. Devroey, B.C. Fauser, Primary ovarian insufficiency, Lancet 376 (9744) (2010) 911–921 (PMID: 20708256).
- [5] E.C. Vogt, F.G. Real, E.S. Husebye, et al., Premature menopause and autoimmune primary ovarian insufficiency in two international multi-center cohorts, Endocr. Connect. 11 (5) (2022) e220024 (PMID: 35521804).
- [6] R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, 2020, CA Cancer J. Clin. 70 (1) (2020) 7–30 (PMID: 31912902).
- [7] R.A. Anderson, F. Amant, D. Braat, et al., ESHRE guideline: female fertility preservation, Hum Reprod Open. 2020 (4) (2020) a52 (PMID: 33225079).
- [8] ASRM, Fertility preservation in patients undergoing gonadotoxic therapy or gonadectomy: a committee opinion, Fertil. Steril. 112 (6) (2019) 1022–1033 (PMID: 31843073).
- [9] Ocebm. Oxford Centre for Evidence-Based Medicine: Levels of Evidence (March 2009) [Z]. 2009.
- [10] Beijing obstetrics and gynecology hospital, Capital Medical University, Committee of Fertility Protection and Preservation of China Association for the Promotion of health science and technology. Specification for ovarian tissue cryopreservation and transplantation. Chinese, Gen. Pract. 26 (23) (2023) 2836–2841 (In Chinese).
- [11] X. Ruan, Chinese Society of Gynecological Endocrinology affiliated to the International Society of Gynecological Endocrinology Guideline for ovarian tissue cryopreservation and transplantation, Gynecol. Endocrinol. 34 (12) (2018) 1005–1010 (PMID: 30129788).
- [12] M.M. Dolmans, M. von Wolff, C. Poirot, et al., Transplantation of cryopreserved ovarian tissue in a series of 285 women: a review of five leading European centers, Fertil. Steril. 115 (5) (2021) 1102–1115 (PMID: 33933173).
- [13] Karavani G, Vedder K, Gutman-Ido E, et al. Prior exposure to chemotherapy does not reduce the in vitro maturation potential of oocytes obtained from ovarian cortex in cancer patients. Hum. Reprod. 2023. doi https://doi.org/10.1093/humre p/dead142. Online ahead of print. PMID: 37414543.
- [14] K.S. Corkum, M.M. Laronda, E.E. Rowell, A review of reported surgical techniques in fertility preservation for prepubertal and adolescent females facing a fertility threatening diagnosis or treatment, Am. J. Surg. 214 (4) (2017) 695–700 (PMID: 28683892).

- [15] M.W. Beckmann, R. Dittrich, L. Lotz, et al., Fertility preservation: complications of surgery and results of removal and transplantation of ovarian tissue, Reprod. Biomed. Online 36 (2) (2018) 188–196 (PMID: 29198423).
- [16] X. Ruan, J. Cheng, J. Du, et al., Analysis of fertility preservation by ovarian tissue cryopreservation in pediatric children in China, Front. Endocrinol. 13 (2022) 930786 (PMID: 35846295).
- [17] J. Cheng, X. Ruan, J. Du, et al., Ovarian tissue cryopreservation for a 3-year-old girl with mosaic turner syndrome in China: first case report and literature review, Front Endocrinol (Lausanne). 13 (2022) 959912 (PMID: 36479213).
- [18] E. Asadi, A. Najafi, J.D. Benson, Comparison of liquid nitrogen-free slow freezing protocols toward enabling a practical option for centralized cryobanking of ovarian tissue, Cryobiology (114) (Dec 12 2023) 104836 (PMID: 38092234).
- [19] H. Khattak, R. Malhas, L. Craciunas, et al., Fresh and cryopreserved ovarian tissue transplantation for preserving reproductive and endocrine function: a systematic review and individual patient data meta-analysis, Hum. Reprod. Update 28 (3) (2022) 400–416 (PMID: 35199164).
- [20] S.E. Gellert, S.E. Pors, S.G. Kristensen, et al., Transplantation of frozen-thawed ovarian tissue: an update on worldwide activity published in peer-reviewed papers and on the Danish cohort, J. Assist. Reprod. Genet. 35 (4) (2018) 561–570 (PMID: 29497953).
- [21] M.M. Dolmans, Y. Iwahara, J. Donnez, et al., Evaluation of minimal disseminated disease in cryopreserved ovarian tissue from bone and soft tissue sarcoma patients, Hum. Reprod. 31 (10) (2016) 2292–2302 (PMID: 27591237).
- [22] M. Shapira, H. Raanani, I. Barshack, et al., First delivery in a leukemia survivor after transplantation of cryopreserved ovarian tissue, evaluated for leukemia cells contamination, Fertil. Steril. 109 (1) (2018) 48–53 (PMID: 29198847).
- [23] M. Nurmio, B. Asadi-Azarbaijani, M. Hou, et al., Effect of previous alkylating agent exposure on follicle numbers in cryopreserved Prepubertal and young adult ovarian tissue after long-term Xenografting, Cancers (Basel). 14 (2) (2022) (PMID: 35053561).
- [24] F. Chevillon, E. Clappier, C. Arfeuille, et al., Minimal residual disease quantification in ovarian tissue collected from patients in complete remission of acute leukemia, Blood 137 (12) (2021) 1697–1701 (PMID: 33171484).
- [25] T. Zver, S. Frontczak, C. Poirot, et al., Minimal residual disease detection by multicolor flow cytometry in cryopreserved ovarian tissue from leukemia patients, J Ovarian Res. 15 (1) (2022) 9 (PMID: 35042558).
- [26] R. Shirai, T. Osumi, D. Keino, et al., Minimal residual disease detection by mutation-specific droplet digital PCR for leukemia/lymphoma, Int. J. Hematol. 117 (6) (2023) 910–918 (PMID: 36867356).
- [27] M.M. Dolmans, J. Donnez, L. Cacciottola, Fertility preservation: the challenge of freezing and transplanting ovarian tissue, Trends Mol. Med. 27 (8) (2021) 777–791 (PMID: 33309205).
- [28] J. Cadenas, P.L. la Cour, L.S. Mamsen, et al., Future potential of IVM including fertility preservation, Fertil. Steril. 119 (4) (2023) 550–559 (PMID: 36702341).
- [29] Practice Committees of the American Society for Reproductive Medicine, the Society of Reproductive Biologists and Technologists, and the Society for Assisted Reproductive Technology. In vitro maturation: a committee opinion. Fertil. Steril. 2021;115(2):298–304. (PMID: 33358333).
- [30] S.C. Braam, V. Ho, T.D. Pham, et al., In-vitro maturation versus IVF: a costeffectiveness analysis, Reprod. Biomed. Online 42 (1) (2021) 143–149 (PMID: 33132059).
- [31] S.J. Silber, S. Goldsmith, L. Castleman, et al., In-vitro maturation and transplantation of cryopreserved ovary tissue: understanding ovarian longevity, Reprod. Biomed. Online 44 (3) (2022) 504–514 (PMID: 35151573).
- [32] X. Ruan, J. Cheng, M. Korell, et al., Ovarian tissue cryopreservation and transplantation prevents iatrogenic premature ovarian insufficiency: first 10 cases in China, Climacteric 23 (6) (2020) 574–580 (PMID: 32508143).
- [33] F.X. Liu, Y. Sun, Identification of the active ingredients and pharmacological effects of Kuntai capsules in the treatment of primary ovarian insufficiency: a review, Medicine (Baltimore) 102 (21) (2023) e33884 (PMID: 37233423).
- [34] J. Byrne, D. Grabow, H. Campbell, et al., PanCareLIFE: the scientific basis for a European project to improve long-term care regarding fertility, ototoxicity and health-related quality of life after cancer occurring among children and adolescents, Eur. J. Cancer 103 (2018) 227–237 (PMID: 30273888).
- [35] C. Poirot, F. Abirached, M. Prades, et al., Induction of puberty by autograft of cryopreserved ovarian tissue, Lancet 379 (9815) (2012) 588 (PMID: 22325664).
- [36] E. Ernst, M. Kjaersgaard, N.H. Birkebaek, et al., Case report: stimulation of puberty in a girl with chemo- and radiation therapy induced ovarian failure by transplantation of a small part of her frozen/thawed ovarian tissue, Eur. J. Cancer 49 (4) (2013) 911–914 (PMID: 23084082).
- [37] K.A. Rodriguez-Wallberg, M. Milenkovic, K. Papaikonomou, et al., Successful pregnancies after transplantation of ovarian tissue retrieved and cryopreserved at time of childhood acute lymphoblastic leukemia - a case report, Haematologica 106 (10) (2021) 2783–2787 (PMID: 34233451).
- [38] C. Poirot, L. Brugieres, K. Yakouben, et al., Ovarian tissue cryopreservation for fertility preservation in 418 girls and adolescents up to 15 years of age facing highly gonadotoxic treatment. Twenty years of experience at a single center, Acta Obstet. Gynecol. Scand. 98 (5) (2019) 630–637 (PMID: 30919447).
- [39] M.M. Dolmans, C. Hossay, T. Nguyen, et al., Fertility preservation: how to preserve ovarian function in children, adolescents and adults, J. Clin. Med. 10 (22) (2021) 5247 (PMID: 34830528).
- [40] I. Demeestere, P. Simon, L. Dedeken, et al., Live birth after autograft of ovarian tissue cryopreserved during childhood, Hum. Reprod. 30 (9) (2015) 2107–2109 (PMID: 26062556).
- [41] S.J. Matthews, H. Picton, E. Ernst, et al., Successful pregnancy in a woman previously suffering from beta-thalassemia following transplantation of ovarian tissue

Practice guidelines

cryopreserved before puberty, Minerva Ginecol. 70 (4) (2018) 432–435 (PMID: 29696941).

- [42] L.B. Colmorn, A.T. Pedersen, E.C. Larsen, et al., Reproductive and endocrine outcomes in a cohort of Danish women following auto-transplantation of frozen/ thawed ovarian tissue from a single center, Cancers (Basel). 14 (23) (2022) (PMID: 36497354).
- [43] X. Ruan, J. Du, D. Lu, et al., First pregnancy in China after ovarian tissue transplantation to prevent premature ovarian insufficiency, Climacteric 24 (6) (2021) 624–628 (PMID: 34374311).
- [44] X. Ruan, J. Du, D. Lu, et al., First live birth in China after cryopreserved ovarian tissue transplantation to prevent premature ovarian insufficiency, Climacteric 25 (4) (2022) 421–424 (PMID: 35504301).
- [45] L. Lotz, J. Bender-Liebenthron, R. Dittrich, et al., Determinants of transplantation success with cryopreserved ovarian tissue: data from 196 women of the Ferti-PROTEKT network, Hum. Reprod. 37 (12) (2022) 2787–2796 (PMID: 36272106).
- [46] F. Jin, X. Ruan, J. Du, et al., Analysis on the characteristics of the patients and effects of ovarian tissue cryopreservation in the first ovarian tissues cryopreservation bank in China, Journal of Capital Medical University. 42 (2021) 521–525 (In Chinese).
- [47] R. Dittrich, J. Hackl, L. Lotz, et al., Pregnancies and live births after 20 transplantations of cryopreserved ovarian tissue in a single center, Fertil. Steril. 103 (2) (2015) 462–468 (PMID: 25487750).
- [48] S. Khiat, P. Bottin, J. Saias-Magnan, et al., Fertility preservation strategies for rectal cancer in reproductive-age women, Future Oncol. 15 (22) (2019) 2635–2643.
- [49] H.S. Taylor, A.M. Kotlyar, V.A. Flores, Endometriosis is a chronic systemic disease clinical challenges and novel innovations, Lancet 397 (10276) (2021) 839–852 (PMID: 33640070).
- [50] J. Donnez, J. Squifflet, M.M. Dolmans, et al., Orthotopic transplantation of fresh ovarian cortex: a report of two cases, Fertil. Steril. 84 (4) (2005) 1018 (PMID: 16213862).
- [51] N. Sanger, M. Menabrito, S.A. Di Spiezo, et al., Fertility preservation counselling for women with endometriosis: a European online survey, Arch. Gynecol. Obstet. 307 (1) (2022) 73–85 (PMID: 35829767).
- [52] G. Calagna, C.L. Della, P. Giampaolino, et al., Endometriosis and strategies of fertility preservation: a systematic review of the literature, Eur. J. Obstet. Gynecol. Reprod. Biol. 254 (2020) 218–225 (PMID: 33011504).
- [53] K. Oktay, O. Oktem, Ovarian cryopreservation and transplantation for fertility preservation for medical indications: report of an ongoing experience, Fertil. Steril. 93 (3) (2010) 762–768 (PMID: 19013568).
- [54] A. Schallmoser, R. Einenkel, C. Farber, et al., Cryostorage of human ovarian tissue: evaluating the storage and disposal pattern over a 22-year period in 2475 patients, Reprod. Biomed. Online 103239 (2023) (PMID: 37400319).
- [55] R. Fabbri, R. Vicenti, V. Magnani, et al., Cryopreservation of ovarian tissue in breast cancer patients: 10 years of experience, Future Oncol. 8 (12) (2012) 1613–1619 (PMID: 23231523).
- [56] J. Donnez, M.M. Dolmans, Fertility preservation in women, N. Engl. J. Med. 377 (17) (2017) 1657–1665 (PMID: 29069558).
- [57] M. von Wolff, N. Sanger, J. Liebenthron, Is ovarian tissue cryopreservation and transplantation still experimental? It is a matter of female age and type of Cancer, J. Clin. Oncol. 01800425 (2018) (PMID: 30289731).
- [58] M. Lambertini, M.L. Del, M.C. Pescio, et al., Cancer and fertility preservation: international recommendations from an expert meeting, BMC Med. 14 (2016) 1 (PMID: 26728489).
- [59] N.D. Ulrich, N.S. Raja, M.B. Moravek, A review of fertility preservation in patients with breast cancer, Best Pract. Res. Clin. Obstet. Gynaecol. 82 (2022) 60–68 (PMID: 35120831).
- [60] X. Ruan, Expert consensus on fertility preservation in hematopoietic stem cell transplantation in girls in China, Gynecol. Endocrinol. 39 (1) (2022) 2146671 (PMID: 36403607).
- [61] E. Biasin, F. Salvagno, M. Berger, et al., Ovarian tissue cryopreservation in girls undergoing haematopoietic stem cell transplant: experience of a single Centre, Bone Marrow Transplant. 50 (9) (2015) 1206–1211 (PMID: 25961773).
- [62] M. Torella, G. Riemma, P. De Franciscis, et al., Serum anti-Mullerian hormone levels and risk of premature ovarian insufficiency in female childhood Cancer survivors, Systematic Review and Network Meta-Analysis. Cancers (Basel). 13 (24) (2021) (PMID: 34944951).
- [63] I. Wikander, F.E. Lundberg, H. Nilsson, et al., A prospective study on fertility preservation in Prepubertal and adolescent girls undergoing hematological stem cell transplantation, Front. Oncol. 11 (2021) 692834 (PMID: 34277437).
- [64] K. Kyono, T. Hashimoto, M. Toya, et al., A transportation network for human ovarian tissue is indispensable to success for fertility preservation, J. Assist. Reprod. Genet. 34 (11) (2017) 1469–1474 (PMID: 28866830).
- [65] N. Ozimek, M. Salama, T.K. Woodruff, National oncofertility registries around the globe: a pilot survey, Front Endocrinol (Lausanne). 14 (2023) 1148314 (PMID: 37223027).
- [66] M. von Wolff, R. Dittrich, J. Liebenthron, et al., Fertility-preservation counseling and treatment for medical reasons: data from a multinational network of over 5000 women, Reprod. Biomed. Online 31 (5) (2015) 605–612 (PMID: 26380870).
- [67] L.M. Ataman, J.K. Rodrigues, R.M. Marinho, et al., Creating a global Community of Practice for Oncofertility, J Glob Oncol. 2 (2) (2016) 83–96 (PMID: 27284576).
 [68] T.K. Woodruff, The Oncofertility consortium-addressing fertility in young people
- with cancer, Nat. Rev. Clin. Oncol. 7 (8) (2010) 466–475 (PMD: 20498666). [69] J. Liebenthron, M. Montag, J. Reinsberg, et al., Overnight ovarian tissue trans-
- portation for centralized cryobanking: a feasible option, Reprod. Biomed. Online 38 (5) (2019) 740–749 (PMID: 30733076).

- [70] E. Isachenko, V. Isachenko, F. Nawroth, et al., Effect of long-term exposure at suprazero temperatures on activity and viability of human ovarian cortex, Fertil. Steril. 91 (4 Suppl) (2009) 1556–1559 (PMID: 19022429).
- [71] J. Liebenthron, M. Montag, Chapter 15 development of a Nationwide network for ovarian tissue cryopreservation, Methods Mol. Biol. 1568 (2017) 205–220 (PMID: 28421499).

Xiangyan Ruan^{a,b,*}, Che Xu^{a,c}, Hefeng Huang^d, Binghe Xu^e, Juan Du^a, Jiaojiao Cheng^a, Fengyu Jin^a, Muqing Gu^a, Weimin Kong^a, Chenghong Yin^a, Yurui Wu^f, Qinjie Tian^g, Yunxia Cao^h, Ruifang Wuⁱ, Liangzhi Xu^J, Jing Jin^a, Yanglu Li^a, Yinmei Dai^a, Rui Ju^a, Fei Ma^e, Gang Wang^k, Wei Wei^a, Xiaojun Huang^J, Maoquan Qin^m, Yuan Linⁿ, Yuan Sun^o, Rong Liu^f, Wei Zhang^p, Xiaodong Li^q, Lin Zou^r, Min Hao^s, Xiyang Ye^t, Fuling Wang^u, Yue Wang^v, Zhuoying Hu^w, Yanhong Huang^x, Tianyuan Zhu^y, Caihong Yang^z, Jinping Wang^{aa}, Xiaomin Yang^{ab}, Rong Ni^{ac}, Liqun Wang^{ad}, Guangxia Luo^{ae}, Aiping Min^{af}, Siyou Zhang^{ag}, Peiling Li^{ah}, Linghui Cheng^h, Lianfang Li^{ai}

- Quanfang Jin^{aj}, Dongmei Shi^{ak}, Yan Li^{al}, Fangying Ren^{am}, Yanxiang Cheng^{an}, Jumin Niu^{ao}, Ying Tian^{ap}, Alfred O. Mueck^{a,b}
- ^a Beijing Obstetrics and Gynecology Hospital, Capital Medical University, Beijing Maternal and Child Health Care Hospital, Beijing, China
- ^b Department for Women's Health, University Women's Hospital and Research Center for Women's Health, University of Tuebingen, Tuebingen, Germany
 - ^c Fuxing Hospital, Capital Medical University, Beijing, China
- ^d Ministry of Education Key Laboratory of Reproductive Genetics, Shool of Medicine, Zhejiang University, Hangzhou, China
 - ^e National Cancer Center/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China
- ^f Children's Hospital, Capital Institute of Pediatrics, Beijing, China ^g Peking Union Medical College Hospital, Peking Union Medical College/
- Chinese Academy of Medical Sciences, Beijing, China ^h The First Affiliated Hospital of Anhui Medical University, Anhui, China
- ⁱ Peking University Shenzhen Hospital, Shenzhen, China
- ^j West China Second University Hospital, Sichuan University, Chengdu, China
- ^k Sichuan Provincial Maternity and Child Health Care Hospital, Chengdu, China
 - ¹ Peking University People's Hospital, Beijing, China
 - ^m National Center for Children's Health, Hematology Center, Beijing
 - Children's Hospital, Capital Medical University, Beijing, China
- ⁿ Fujian Maternity and Child Health Hospital College of Clinical Medicine for Obstetrics & Gynecology and Pediatrics, Fujian Medical University, Fujian, China
 - ^o Beijing Jingdu Children's Hospital, Beijing, China
- ^p Obstetrics and Gynecology Hospital of Fudan University, Shanghai, China
 - ^q The First Hospital of Hebei Medical University, Hebei, China
- ^r Affiliated Hospital of Guangdong Medical University, Guangdong, China ^s Second Hospital of Shanxi Medical University, Shanxi, China
- ^t Shenzhen People's Hospital (The Second Clinical Medical College, Jinan
- University; The First Affiliated Hospital, Southern University of Science and
 - Technology), Shenzhen, China
 - ^u The Affiliated Hospital of Qingdao University, Qingdao, China
 - ^v Henan Provincial People's Hospital, Zhengzhou, China
- ^w The First Affiliated Hospital of Chongqing Medical University, Chongqing, China
 - ^x Xi'an International Medical Center Hospital, Xi'an, China
 - ^y Gansu Provincial Maternal and Child-care Hospital/Gansu Province Central Hospital, Lanzhou, China
 - ^z The General Hospital of Ningxia Medical University, Ningxia, China ^{aa} Zibo Maternal And Child Health Hospital, Zibo, China
 - ^{ab} Liuzhou Maternity and Child Healthcare Hospital, Liuzhou, China
- ^{ac} The Central Hospital of Enshi Tu Jia and Miao Autonomous Prefecture, Enshi, China
 - ^{ad} Jiangxi Maternal and Child Health Hospital, Jiangxi, China

^{ae} The First People's Hospital of Huaihua (Hunan University of Medicine General Hospital), Huaihua, China

Practice guidelines

^{an} Renmin Hospital of Wuhan University, Wuhan, China ^{ao} Shenyang Women's and Children's Hospital, Shenyang, China ^{ap} XiangXi Ninger Obstetrics and Gynecology Hospital, Xiangxi, China

^{*} Corresponding author at: Department of Gynecological Endocrinology, Beijing Obstetrics and Gynecology Hospital, Capital Medical University, Beijing Maternal and Child Health Care Hospital, No. 251, Yaojiayuan Road, Chaoyang District, Beijing 100026, China. *E-mail address:* ruanxiangyan@ccmu.edu.cn (X. Ruan).

^{af} People's Hospital of Leshan City, Leshan, China

^{ag} The First People's Hospital of Foshan, Foshan, China ^{ah} The Second Affiliated Hospital of Harbin Medical University, Harbin,

China

^{ai} Beijing Obstetrics and Gynecology Hospital, Capital Medical University, Huairou Maternal and Child Health Care Hospital, Huairou, China

^{aj} Shanghai Baoshan Hospital of Integrated Traditional Chinese and Western Medicine, Shanghai, China

^{ak} Maternal and Child Health Hospital of Yinchuan, Yinchuan, China ^{al} Luoyang Anhe Hospital, Luoyang, China

am People's Hospital of Linxi County, Hebei, China