DOI: 10.1111/jdv.20055

ORIGINAL ARTICLE



JEADV

International Skin Imaging Collaboration-Designated Diagnoses (ISIC-DX): Consensus terminology for lesion diagnostic labeling

Alon Scope^{1,2} | Konstantinos Liopyris^{2,3} | Jochen Weber² | Raymond L. Barnhill⁴ | Ralph P. Braun⁵ | Clara N. Curiel-Lewandrowski⁶ | David E. Elder⁷ | Gerardo Ferrara⁸ | Jane M. Grant-Kels^{9,10} | Thiago Jeunon¹¹ | Aimilios Lallas¹² | Jennifer Y. Lin¹³ | Michael A. Marchetti² | Ashfaq A. Marghoob² | Cristian Navarrete-Dechent¹⁴ | Giovanni Pellacani¹⁵ | Hans Peter Soyer¹⁶ | Alexander Stratigos¹⁷ | Luc Thomas¹⁸ | Harald Kittler¹⁹ | Veronica Rotemberg² | Allan C. Halpern²

Correspondence

Alon Scope, The Kittner Skin Cancer Screening & Research Institute, Sheba Medical Center, Ramat Gan 5262000, Israel. Email: scopea1@gmail.com

Funding information

Israel Science Foundation, Grant/Award Number: ISF-1546-16; National Cancer Institute, Grant/Award Number: P30-CA008748; National Health and Medical Research Council, Grant/Award Number: APP1137127

Abstract

Background: A common terminology for diagnosis is critically important for clinical communication, education, research and artificial intelligence. Prevailing lexicons are limited in fully representing skin neoplasms.

Objectives: To achieve expert consensus on diagnostic terms for skin neoplasms and their hierarchical mapping.

Methods: Diagnostic terms were extracted from textbooks, publications and extant diagnostic codes. Terms were hierarchically mapped to super-categories (e.g. 'benign') and cellular/tissue-differentiation categories (e.g. 'melanocytic'), and appended with pertinent-modifiers and synonyms. These terms were evaluated using a modified-Delphi consensus approach. Experts from the International-Skin-Imaging-Collaboration (ISIC) were surveyed on agreement with terms and their hierarchical mapping; they could suggest modifying, deleting or adding terms. Consensus threshold was >75% for the initial rounds and >50% for the final round.

Results: Eighteen experts completed all Delphi rounds. Of 379 terms, 356 (94%) reached consensus in round one. Eleven of 226 (5%) benign-category terms, 6/140 (4%) malignant-category terms and 6/13 (46%) indeterminate-category terms did not reach initial agreement. Following three rounds, final consensus consisted of 362 terms mapped to 3 super-categories and 41 cellular/tissue-differentiation categories. **Conclusions:** We have created, agreed upon, and made public a taxonomy for skin neoplasms and their hierarchical mapping. Further study will be needed to evaluate the utility and completeness of the lexicon.

INTRODUCTION

Clinical care requires clear communication between clinicians and pathologists about diagnostic categories that will guide management.¹ To be useful for educational, research and artificial intelligence (AI) training purposes, large skin imaging data sets need agreed-upon diagnostic labels with a clear-cut hierarchy.^{2–8}

The International Skin Imaging Collaboration (ISIC) was established to facilitate use of skin imaging towards reducing mortality and morbidity from skin cancer.⁸ A goal of ISIC is to standardize image acquisition, diagnostic labeling, metadata terminology and digital archiving towards

For affiliations refer to page 123.

J Eur Acad Dermatol Venereol. 2025;39:117–125.

Linked article: M. Suppa et al. J Eur Acad Dermatol Venereol 2025;39:33-34. https://doi.org/10.1111/jdv.20438.

^{© 2024} European Academy of Dermatology and Venereology.

development of a large image data set for educational and research purposes. Indeed, the ISIC image archive is a growing, open-source repository of skin neoplasms, available for teaching, conducting research and developing AI systems.⁹ The ISIC archive has been the source of dermoscopic images for five international grand challenges leading to thousands of technical publications.^{4,10,11} To date, ISIC has restricted itself to a dozen diagnostic categories of skin neoplasms. As ISIC archive broadens to represent the full gamut of skin proliferations, we find that prevailing dermatological lexicons are limited in their scope, consistency and granularity.

Herein, we focused on trying to achieve standardized diagnostic labeling of images of skin neoplasms based on correspondence with histopathological terminology. To this end, we have conducted a modified Delphi-consensus predicated on existing diagnostic lexicons and present the resulting taxonomy for broad international use.

MATERIALS AND METHODS

Consistent with previous studies, an institutional review board approval was not applicable to this study, as it included no research subjects¹²; all Delphi members were co-authors. Illustrative clinical images were retrospectively selected from an image database; written informed consent to publication of their case details was obtained from the patients whose images are depicted.

Study aims and initial development of diagnostic scheme

Our plan is to label existing and future ISIC-Archive images with diagnostic terms that most accurately and reproducibly represent the histopathological diagnoses. For each clinicalpathological entity, we sought the best diagnostic term, the pertinent modifiers and commonly used synonyms and the hierarchical mapping of diagnoses.

We began with several premises: (1) focus on neoplasms with few examples of inflammatory processes that may appear in the clinical differential diagnosis of skin lesions, and (2) convergence on agreed-upon diagnostic labels—prioritizing a unique ISIC-designated-diagnosis (ISIC-DX) term for each clinical-pathological entity.

Next, we extracted diagnostic terms from dermatopathology textbooks, scientific literature, online sources and extant diagnostic codes.^{13–17} We created a three-tier hierarchy with the top level mapped to a 'benign' (Figure 1), 'malignant' (Figure 2) or 'indeterminate' (Figure 3) super-category.^{18–20} Next, each diagnosis was mapped to cellular/tissue differentiation category; these categories were termed based on the cell type (e.g. 'melanocytic') or on the tissue element (e.g. 'vascular' or 'neural') from which the proliferation arises or with which it is closely associated. Synonyms were added to terms, to recognize variable diagnostic designations commonly used for the same entity. Finally, modifiers were

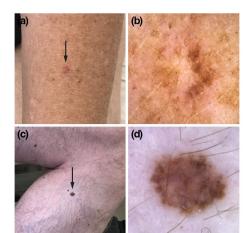


FIGURE 1 Examples of benign proliferations. (a) Clinical and (b) dermoscopic images of a lesion on the leg histopathologically diagnosed as 'dermatofibroma'. This lesion is classified into super-category 'Benign', cellular/tissue differentiation category. This lesion is classified into super-category 'Benign', cellular/tissue differentiation category 'Soft tissue proliferations – Fibro-histiocytic', diagnostic term 'Dermatofibroma, NOS. (c) Clinical and (d) dermoscopic images of a lesion on the leg histopathologically diagnosed as 'dysplastic compound nevus'. It is classified into super-category 'Benign', cellular/tissue differentiation category 'Melanocytic proliferations', diagnostic term 'Nevus, Atypical, NOS/Nevus, Dysplastic/Nevus, Clark', with modifier 'Compound'.

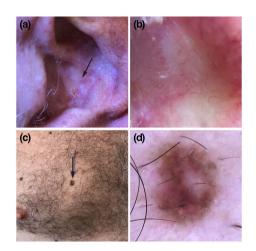


FIGURE 2 Examples of malignant proliferations. (a) Clinical and (b) dermoscopic images of a lesion on the ear concha histopathologically diagnosed as 'Basal cell carcinoma, sclerosing (morphea) type'. This lesion is classified into super-category 'Malignant', cellular/tissue differentiation category 'Adnexal epithelial proliferations – Follicular', diagnostic term 'Basal cell carcinoma, Sclerosing/morpheaform'. (c) Clinical and (d) dermoscopic images of a lesion on the chest histopathologically diagnosed as 'melanoma 0.5 mm in thickness, without ulceration, and with nevoid features'. It is classified into super-category 'Malignant', cellular/tissue differentiation category 'Melanocytic proliferations (melanoma)', diagnostic term 'Melanoma invasive, Nevoid', with modifiers 'Breslow depth 0.5 mm' and 'non-ulcerated'.

defined as descriptors that allow further sub-categorization of terms, in a manner that is clinically or pathologically meaningful. For example, nevi are often sub-classified by pathologists as 'junc-tional', 'compound' or 'intradermal'.

Delphi contributors and rounds

We utilized a modified Delphi method to attain consensus on the terminology lexicon.^{21,22} We invited members of ISIC who are dermatologists and dermatopathologists to serve as Delphi experts by sending an email describing the study aims and Delphi methodology and asking recipients to preview the study user interface via a 6.5-min tutorial video.²³ Consent to serve as experts in the Delphi process was implied through self-registration and round completion.

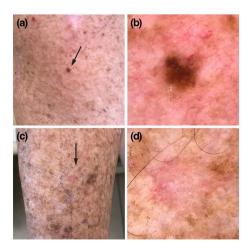


FIGURE 3 Examples of Indeterminate proliferations. (a) Clinical and (b) dermoscopic images of a lesion on the arm histopathologically diagnosed as 'atypical junctional melanocytic lesion, in favor of melanoma in-situ'. This lesion is classified as super-category 'Indeterminate', cellular/ tissue differentiation category 'Melanocytic proliferations', diagnostic term 'Atypical melanocytic neoplasm' with modifiers 'Junctional', 'MPATH III' and 'Histopathological report favours melanoma'. (c) Clinical and (d) dermoscopic images of a lesion on the forearm, whereby the histopathological report described an 'atypical junctional melanocytic hyperplasia on sun-damaged skin, the possibility of a developing melanoma in situ cannot be excluded'. It is classified into super-category 'Indeterminate', cellular/tissue differentiation category 'Melanocytic proliferations', diagnostic term 'Atypical melanocytic neoplasm' with modifiers 'Junctional', 'MPATH III' and 'Histopathological report 'Melanocytic proliferations', diagnostic term 'Atypical melanocytic neoplasm' with modifiers 'Junctional', 'MPATH III' and 'Histopathological report undecided'.

Contributors were surveyed on basic demographics and could recommend inclusion of additional experts as Delphi members. Contributors who completed all Delphi rounds were offered authorship in the ensuing publication.

In the first Delphi round, contributors were asked to review the entire list of ISIC-DX terms using a web interface (Figure 4). Contributors had to respond to each individual ISIC-DX term by marking the term as 'correct', denoting agreement with the term, its hierarchical mapping and when relevant, its synonyms and modifiers. Alternatively, they could select a modification: (1) suggest a different name for a term or category, (2) correct spelling of the term, (3) combine with another term, (4) assign term to another category in the hierarchical mapping, (5) edit synonyms or modifiers, (6) suggest additional synonyms or modifiers, and (9) other free text suggestions. In addition, contributors could (1) suggest adding new categories in the hierarchical mapping, and (2) suggest adding new terms. At all times, they could refer to an overview of the entire taxonomy.

Results of the first round were evaluated after all experts submitted their surveys. Threshold for Delphi consensus was set at >75%.²⁴ A steering committee of three ISIC members (AS, VR and KL) who were not in the Delphi panel reviewed contributors' responses for terms that did not reach consensus and clustered these responses to multiple-choice items. In addition, the committee made majority-vote decisions on suggestions for added terms. These items formed the basis for the second Delphi round survey. Suggestions for new terms were not accepted after round one. At the second round, consensus threshold was retained at >75% for multiple-choice (>2) items, while threshold was set at a majority vote of >50% for items with only two choices. For items that did not reach consensus, we identified the top-two voted choices, for final evaluation at the third Delphi round. At the conclusion of the third round, threshold for consensus was set at a majority vote of >50%. For third round items with tied participant votes, the steering committee made a majority-vote decision. Delphi surveys and participants responses are available in a public repository.²⁵

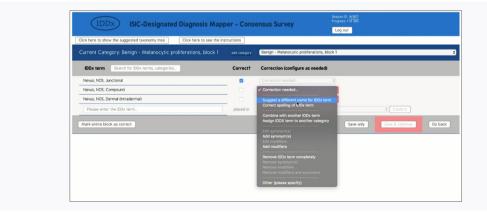


FIGURE 4 Screenshot of the web interface presented to members in the first Delphi round evaluation. In this exemplary page, Delphi members were presented with a batch of three ISIC-DX terms, Nevus, NOS-junctional, -compound and -dermal. The upper right box shows the hierarchical mapping of the terms to 'benign' super-category and 'melanocytic proliferations' category. Checking the 'correct?' box denoted agreement with the term. Alternatively, they could use a drop-down menu to suggest deleting or modifying the term or its associated mapping.

Statistical design

Descriptive and relative frequencies were used to describe the survey respondents and the results of the consensus. Measures of central tendency were calculated. Agreement threshold >75% for 18 Delphi members was calculated as \geq 14 consensus votes and >50% as \geq 10 votes.

RESULTS

Delphi contributors

Invitations to contribute to the study were sent to 36 clinicians. Of 36 invited, 18 (50%) completed all Delphi rounds (10 dermatologists, four dermato-pathologists and four with a dual certification); five (14%) initially agreed to be Delphi members, but did not complete round I; three (8%) declined and 10 (28%) did not respond.

malignant = 140 and indeterminate = 13). Of 379 terms, 356 (94%) passed the 75% threshold in round I. Twenty-three terms, including 11/226 (5%) benign-category terms, 6/140 (4%) malignant-category terms and 6/13 (46%) indeterminate-category terms, did not reach agreement. These were carried over to round II as 34 items, including ISIC-DX terms (n=16), modifiers (n=7), synonyms (n=8), assignment of terms to another category in the hierarchical mapping (n=2) and change of cellular/tissue differentiation category name in the hierarchical mapping (n=1). In addition, 29 newly suggested terms were introduced into round II.

Based on the contributor feedback, the steering committee decided to remove the inflammatory (n = 15) and infectious terms (n = 12) and focus solely on skin neoplasms. This reduced the total number of approved ISIC-DX terms, at the conclusion of round I, from 356 to 329 terms.

Round II

Round I

Round I was held between November 2019 and January 2020. Overall, there were 379 ISIC-DX terms (benign=226,

Round II was held from August to October 2020. Contributors voted on 63 items carried over from round I. Of 34 items related to existing terms, 22 items (65%) surpassed the agreement threshold, including 6/16 (38%) terms, 7/7 (100%) modifiers, 7/8 (88%) synonyms, 1/2 (50%) for assignment of

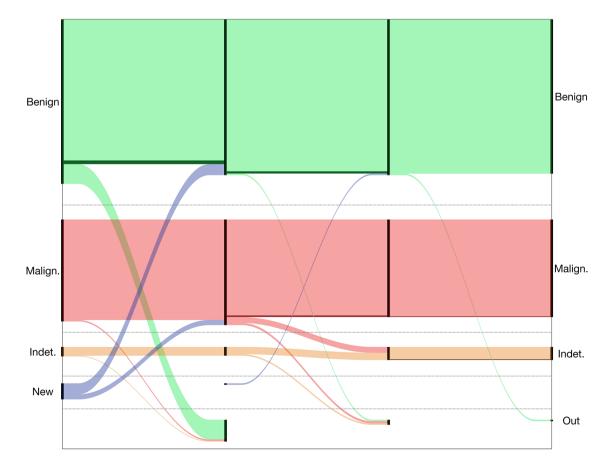


FIGURE 5 Alluvial plot depicting the amendments made in the ISIC-DX taxonomy across the three rounds of Delphi. For example, the green thick curved line shows the fraction of terms from the benign super-category that were eliminated after round I. The two purple thick curved lines on the left shows new terms introduced into the benign (top line) and malignant (bottom line) super-categories following round I.

terms to another category in the hierarchical mapping and 1/1 (100%) for change of cellular/tissue differentiation category name in the hierarchical mapping-under 'benign' super-category, 'deposits' was renamed 'exogenous'. In addition, 26 of the 29 (90%) new terms introduced in round II exceeded the agreement threshold. Fourteen multiple-choice items did not reach the agreement threshold; the two terms with the highest number of votes for each item were carried over to Delphi round III. Finally, one binary-choice itemshould 'solar lentigo' be categorized under 'flat melanotic pigmentations' or under 'epidermal proliferations'-had a tie of votes (9/18 for each option); the steering committee made a majority vote to retain the original classification of 'solar lentigo' under 'epidermal proliferations', even though some solar lentigines exhibit a measure of melanocytic hyperplasia.

Round III

Round III was held between February and April 2021. Contributors voted on 14 entities from round II, including existing terms (n=10), newly suggested terms (n=2), synonyms (n=1) and assignment of terms to a category (n=1). Of these, 13 items (93%) reached consensus by majority vote. 4683083, 2025, 1, Downloaded from https://onlinelibrary.wiley.com/doi/10.1111/jdv.20055 by CAPES, Wiley Online Library on [15/01/2025]. See the Terms and Conditions (https://onlinelibrary.wiley.com/terms

and-conditions) on Wiley Online Library for rules of use; OA articles

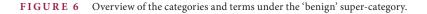
are governed by the applicable Creative Commons License

There was one item—whether to combine 'Nevus, Atypical, NOS', 'Nevus, Dysplastic' and 'Nevus, Clark' into one ISIC-DX, or to Combine 'Nevus, Atypical, NOS' and 'Nevus, Dysplastic', while deleting 'Nevus, Clark'—that did not reach consensus because of a tied vote. The steering committee made a majority vote for the former option of combining all three into one term, namely 'Nevus, Atypical, NOS/ Nevus, Dysplastic/Nevus, Clark', classified under 'benign' 'melanocytic proliferations'.

In addition, the steering committee added two survey items to maintain consistency of the terminology. First, contributors decided in round II to move the term 'solar (actinic) cheilitis' from 'malignant' to 'indeterminate' super-category. In round III, they were surveyed and confirmed that all 'solar keratosis' terms be moved from 'malignant' to 'indeterminate' super-category, under 'epidermal proliferations'. Second, contributors decided in round II to delete the term 'PUVA keratoses'. In round III, they were asked whether the term 'arsenical keratoses' should be deleted as well and voted in favour of deletion of this term.

An overview of the amendments made in the ISIC-DX taxonomy across the three rounds of Delphi is depicted (Figure 5). The final consensus list consists of 362 ISIC-DX terms (Appendix S1). The 'benign' super-category encompasses 20 categories and 210 terms (Figures 1 and 6); the

Vielanocytic proliferations	Adnexal epithelial proliferations - Follicular	Soft tissue proliferations - Vascular	Soft tissue proliferations - Adipocytic
Vevus, NOS, Junctional	Trichoblastoma, NOS	Hemangioma, NOS	Lipoma, NOS
levus, NOS, Compound	Trichoepithelioma	Hemangioma, Cherry	Lipoma, Spindle cell / pleomorphic
levus, NOS, Dermal (Intradermal)	Trichoepithelioma, Desmoplastic	Hemangioma, Infantile	Angiolipoma
levus, Atypical, NOS / Nevus, Dysplastic / Nevus, Clark	Tricholemmoma	Noninvoluting congenital hemangioma (NICH)	Fibrolipoma
Vevus, Arypical, NOS / Nevus, Dysplastic / Nevus, Clark	Tricholemmoma, Desmoplastic	Rapidly involuting congenital hemangioma (RICH)	Lipomatous nevus
entiginous melanocytic proliferation	Proliferating tricholemmal tumor	Hemangioma, Hobnail	Benign soft tissue proliferations, Adipocytic, other (to be specified)
levus, Spitz	Folliculosebaceous cystic hamartoma	Hemangioma, Tufted	Soft tissue proliferations - Neural
levus, Reed	Pilomatricoma	Glomeruloid hemangioma	· · · · · · · · · · · · · · · · · · ·
Vevus, BAP-1 deficient	Trichofolliculoma	Acquired elastotic hemangioma	Neuroma, NOS
Vevus, Congenital	Proliferating tricholemmal tumor	Verrucous hemangioma	Neuroma, Traumatic
(per history - present at birth or within 1 year of infancy)	Pilar sheath acanthoma	Hemangioma, other (to be specified)	Neuroma, Palisaded and encapsulated
levus, Congenital pattern (per histopathology)	Nevus comedonicus	Angiokeratoma	Neurofibroma, NOS
Proliferative nodule in congenital melanocytic nevi	Warty dyskeratoma	Pyogenic granuloma	Neurofibroma, Plexiform
(Minimal or no atypia AND no mitotic activity (MPATH II)	Tumor of folicular infundibulum	Glomus tumor	Granular cell tumor
Vevus, Spilus	Panfolliculoma		Granular cell tumor, Non-neural
Vevus, Agminated		Glomangiomyoma	
lue nevus, NOS	Benign adnexal epithelial proliferations, Follicular or Infundibular, other	Angioleiomyoma	Schwannoma
	(to be specified)	Angiolymphoid hyperplasia with eosinophilia	Nerve sheath myxoma
llue nevus, Common		Nevus anemicus	Perineurioma
Rue nevus, Cellular		Capillary vascular malformation	Benign soft tissue proliferations, Neural, other (to be specified)
Blue nevus, Epithelioid		Venous malformation	
Nue nevus, Sclerosing	Adnexal epithelial proliferations - Sebaceous	Arterio-venous malformation	Soft tissue proliferations - Cartilagenous and ossifyir
Nue nevus, Plaque type			Accessory tragus
Vevus, Deep penetrating	Sebaceous hyperplasia	Lymphangioma (superficial lymphatic malformation)	Extraskeletal chondroma
vevus, Deep penetrating Vevus, Combined (details of 2 components to be specified)	Sebaceous adenoma	Other vascular or lymphatic malformation / hamartoma	
	Sebaceoma	(to be specified)	Subungual osteochodroma
levus, Combined, NOS	Nevus sebaceus	Vascular spider	Osteoma cutis
levus, Acral	Fordyce spots	Venous lake	Benign soft tissue proliferations, Cartilagenous and ossifying, other
levus, Of special anatomic site	Fibrofoliculoma	Telangiectasia	(to be specified)
(e.g., genital, flexural, milkline and breast, other)		Acroangiodermatitis of Mali	Soft tissue proliferations - Myoepithelial
Vevus, Recurrent / persistent	Trichodiscoma	Benign soft tissue proliferations, Vascular, other (to be specified)	Sort ussue promerations - wyoepithelia
Vevus, Halo	Benign adnexal epithelial proliferations, Sebaceous, other	benigh sore assue promerations, vascular, other to be specified)	Myoepithelioma
Vevus, Balloon cell	(to be specified)	Soft tissue proliferations - Fibro-histiocytic	
		Soft ussue promerations - horo-histocytic	Mast cell proliferations
Vevus, Meyerson		Dermatofibroma, NOS	
Lentigo simplex		Dermatofibroma, Aneurysmal	Mastocytosis, NOS
Mongolian spot	Adnexal epithelial proliferations - Apocrine or Eccrine	Dermatofibroma, Hemosiderotic	Mastocytoma, Solitary / unifocal
Vevus of Ito	Autexal epithelial proliferations - Apochine of Ecchine		Maculopapular mastocytoma
Vevus of Ota	Syringoma	Dermatofibroma, Epithelioid	Mastocytosis, Diffuse / multifocal (subtype: Telangiectasia mcularis
igmented epithelioid melanocytoma	Poroma	Dermatofibroma, Cellular	eruptiva perstans, Diffuse cutaenous, other; to be specified)
Dermal melanocytosis, NOS	Hidradenoma, Apocrine, NOS	Dermatofibroma, Atypical	Mast cell proliferation, other (to be specified)
		Dermatofibroma, other (to be specified)	
Benign melanocytic proliferations, other (to be specified)	Hidradenoma, Apocrine, Predomnantly with clear cells	Non-Langerhabs histiocytosis, NOS	Langerhans cell proliferations
	Hidradenoma, Poroid	Juvenile xanthogranuloma	
	Hidradenoma papiliferum	Rosai-Dorfman disease	Langerhans cell histiocytosis, NOS
	Hidradenoma, NOS		Langerhans cell histiocytosis, Solitary / unifocal
	Spiradenoma	Reticulohistiocytosis	Langerhans cell histiocytosis, Diffuse / multifocal
Hat melanotic pigmentations (not melanocytic nevus)	Cylindoma	Non-Langerhabs histiocytosis, other (to be specified)	Langerhans cell histiocytosis, other (to be specified)
	Mixed tumor, Apocrine type	Fibroepithelial polyp	Indeterminate cell histiocytosis, NOS
nk-spot lentigo		Angiofibroma, Facial	
Aucosal melanotic macule	Mixed tumor, Eccrine type	Angiofibroma, Penile	Erdheim Chester disease
Café au lait macule / patch	Mixed tumor, NOS	Angiofibroma, Periungual	Mixed Langerhans cell histiocytosis and Erdheim Chester disease
phelis	Syringocystadenoma papilliferum	Myxoma, Cutaneous	Cysts
	Apocrine tubular adenoma	Fibroma, NOS	
	Syringofibroadenoma		Cyst, NOS
	Supernumerary nipple	Fibroma, Sclerotic	Milium
	Fibroadenoma	Fibroma, Pleomorphic	Comedo
Teldenned uneliferations		Fibroma, other (to be specified)	Infundibular / epidermal cyst
Epidermal proliferations	Myoepithelioma	Giant cell turnor of the tendon sheath	Trichilemmal / isthmic-catagen / pilar cyst
iolar lentigo	Cystadenoma	Scar, NOS	
inharthais konstasis	Benign adnexal epithelial proliferations, Apocrine or Eccrine, other	Scar, Hypertrophic	Dilated pore
ieborrheic keratosis ieborrheic keratosis, Clonal	(to be specified)	Scar, Keloid	Steatocystoma
eborrneic keratosis, Cional			Digital mucous cyst
ichen planus like keratosis		Benign soft tissue proliferations, Fibro-histiocytic, other	Benign cyst, other (to be specified)
Clear cell acanthoma		(to be specified)	
arge cell acanthoma	Hemorrhagic lesions (extravasation of erythrocytes)	Coft tissue proliferations . Musels tissue or pro-film-blactic	-Exogenous
Porokeratosis		Soft tissue proliferations - Muscle tissue or myofibroblastic	Tattoo
ipidermal nevus	Hemorrhage, NOS	Dermatomyofibroma	
	Hemorrhage, Subcorneal and intracorneal	Smooth muscle hamartoma	Foreign body granuloma
pidermolytic acanthoma	Hemorrhage, Dermal and subcutaneous		Exogenous, other (to be specified)
Acantholytic acanthoma		Piloleiomyoma	Collision - Only benign proliferations
Melanoacanthoma	Hemorrhage, Subungual	Angioleiomyoma	consion - only benign promerations
Benign epidermal proliferations, other (to be specified)	Hemorrhage, Mucosal	Dartoic muscle leiomyoma	Collision, Only benign proliferations (to be specified)
	Hemorrhage, other (to be specified)	Nodular Fasciitis	Transition and the second of the second of
	nemormage, other (to be specified)		
	nemorrhage, ourier (to be specified)	Benign soft tissue proliferations, Myofibrobastic or muscle tissue,	Benign - Other



'malignant' super-category—19 categories and 134 terms (Figures 2 and 7); and the 'indeterminate' super-category—2 categories, namely 'melanocytic proliferations' and 'epider-mal proliferations', and 18 terms (Figures 3 and 7).

DISCUSSION

Herein, we present a Delphi-based, proposed clinicalpathological categorization of skin neoplasms. We share our initial attempt to develop a consensus lexicon designed for use as diagnostic labels for improved clinician communication, education, research and AI training. We also plan to apply this terminology for labeling of the images used in the ISIC Archive.

Our initial strategy was to adopt an existing authoritative taxonomy, however, we found them lacking in scope, granularity and consistency. There have been efforts to standardize diagnostic terminology, including ICD10²⁶ and SNOMED.²⁷ Yet, the clinical utility and reproducibility of terms used by these coding systems have been previously called into question.^{28–30} For example, among benign melanocytic proliferations, the current Delphi consensus recorded 36 ISIC-DX terms; of these, 26 would not have a distinctive ICD-10 diagnosis, which could detract from the clarity of diagnostic labeling of the ISIC archive. To fill some of these gaps, pathologists have suggested additional structured reporting schemes, such as the MPATH-Dx and the codes of the College of American Pathologists.^{1,31} Indeed, MPATH-Dx provides an encompassing scheme for structured labeling of melanocytic proliferations with associated treatment recommendations.³²⁻³⁴

Ideally, all neoplasms could be categorized into benign versus malignant super-categories. In reality, as highlighted by MPATH-Dx, there are many examples of lesions (e.g. 'Atypical Intraepithelial Melanocytic Proliferation') for which their malignant versus benign status is debated. However, exclusion of such relevant 'borderline' cases from clinical training and education and from AI training risks undermining the diagnostic accuracy of clinicians in daily practice, as well as that of AI systems. Hence, we added an 'indeterminate' super-category to the first tier of our hierarchical scheme. Almost half (46%) of the ISIC-DX terms under the 'indeterminate' super-category did not reach consensus in the first round of the Delphi, while about 95% of the terms under 'benign' and 'malignant' supercategories did. This likely reflects clinicians' ambiguous view of entities whose biological behaviour is still deliberated. While the 'indeterminate' super-category initially only included the category 'melanocytic proliferations', the consensus panel decided to include all terms related to 'solar (actinic) keratosis' under 'indeterminate' 'epidermal proliferations'. This categorization echoes an old literature deliberation on the nature of solar keratosis.^{35–37} However,

Melanocytic proliferations	Epidermal proliferations	Adnexal epithelial proliferations - Apocrine or Eccrine	Soft tissue proliferations - Neural
Melaroma in situ, NOS Melaroma in situ, Lontjon naligna type / melaroma in situ, Lontjon naligna type / Melaroma in situ, Lontjon naligna in the second skin Melaroma in situ, Nacosal Site to be specified e.g. iki, onal mucosa, genital, other) Melaroma in situ, Superficial spreading Melaroma in situ, Recurrent / persistent Melaroma in situ, Becurrent / persistent Melaroma in situ, other (to be specified) Melaroma in situ, other (to be specified) Melaroma in suba, NOS Melaroma invasive, Superficial spreading Melaroma invasive, Superficial spreading Melaroma invasive, NOS Melaroma invasive, NoS On chronicity, Anzi / Acral-lentiginous Melaroma invasive, Moduar Melaroma invasive, Moduar	uamous cel carcinoma in situ, NOS Paget disease, Kran-mammary Paget disease, Extra-mammary Paget disease, Paget di		Soft tissue proliferations - Neural Malgant granulation cell turor Malgant granulation cell turor Malgant soft tissue proliferations, Neural, other (to be specified) Soft tissue proliferations - Cartilagenous and ossifying Extrasledetal osteosarcoma Malgant soft tissue proliferations - Cartilagenous and ossifying, other to be specified Soft tissue proliferations - Myoepithelial Myoepithelial sarcoma Soft tissue proliferations - Unknown or other histiogenesi Avalignmat off tusse proliferation, Unknown or other histiogenesis, other (to be specified) Merkel cell proliferation Merkel cell proliferation
Melanoma Invasive, Neurotropic Melanoma Invasive, Spitzold (melanoma reveniting Spitz neus / tumor) Melanoma Invasive, Spitzold (melanoma reveniting Spitz neus / tumor) Melanoma Invasive, Pigmented spitzle (of neus) like (melanoma reveniting or originating from a blue neus) Melanoma Invasive, Maccold Melanoma Invasive, Maccold Melanoma Invasive, Maccold (resembling optichedi blue neus) Melanoma Invasive, Maccold Melanoma Invasive, Maccold (resembling optichedi blue neus) Melanoma Invasive, Harsing in a congenital neus Melanoma Invasive, Farsing in a congenital neus Melanoma Invasive, ether (to be specified) Melanoma Invasive, other (to be specified) Melanoma metastasis Skin metastasis of internal solid (non-hematological) cancer	Adhexal epithelial proliferations - Folicular Basal cell carcinoma, NOS Basal cell carcinoma, NoS Basal cell carcinoma, Nodraf Basal cell carcinoma, Nordraf Basal cell carcinoma, Nordraf Basal cell carcinoma, Nordraf Basal cell carcinoma, Nordraf Basal cell carcinoma and adresal differentiation Basal cell carcinoma with sarcomaticol differentiation Basal cell carcinoma with sarcomaticol differentiation Basal cell carcinoma, Combined subtypes (lettelis of all components to be specified) Basal cell carcinoma, cother (to be specified) Matrical / plomatrical carcinoma Malginant advalid-gethelial proliferations, Folicular, other (to be specified)	Angiosarcoma cutaneous, NOS Angiosarcoma cutaneous, NOS Angiosarcoma cutaneous, Vith associated lymphedema Angiosarcoma cutaneous, Post-tradistion Angiosarcoma cutaneous, Post-tradistion Maignant giornus turnor Hemangioendorheliona, Alco Los specifical Maignant soft tissue proliferations, Venziar, other (to be specified) Maignant soft tissue proliferations, Venziar, other (to be specified) Soft tissue proliferations – Fibro-histiocytic Asypical finanantoma Bernard Strates and Strates and Strates Permatifications protuberase (DFSP) Firoarcoma Epitheliol discorma Maignant Soft tissue proliferations, Fibro-histiocytic, other (to be specified) Soft tissue proliferations - Muscle tissue or myofibroblastic Leoropaarcoma, Cutaneous	Lymphocytic proliferations - T-Cell/NK Mycosis fungaides, NGS Mycosis fungaides, Foliadoropic Mycosis fungaides, Foliadoropic Mycosis fungaides, Sofiadoropic Mycosis fungaides, Graniumatous Seary syndrome Adult T-cell kymphona Frimary utaneous CD4+ small/medum T-cell kymphona Frimary utaneous CD4+ hymphoproliferative disese, NOS Primary utaneous CD4+ hymphoproliferative disese, Cutanous anaptitic large cell hymphoma Extrandial MycT-ell kymphoma Extrandial MycT-ell kymphoma Extrandial MycT-ell kymphoma Extrandial MycT-ell kymphoma Extrandial MycT-ell kymphoma Extrandial MycT-ell kymphoma Extrandial MycT-ell kymphoma, Rare subtype, NOS Primary utaneous peripheral T-cel kymphoma, Rare subtype (to be specified)
(non-hematological) cancer (to be specified) Collision - At least one malignant proliferation Collision, At least one malignant proliferation (to be specified) Malignant - Other Malignant, other (or not readily classifiable; to be specified)	Adnexal epithelial proliferations - Sebaceous Sebaceous cardnoma Malganat advecal epithelial proliferations, Sebaceous, other (to be specified)	Rhabdomyoscaroma, Cutaneous Apycial Intrademul Smooth muscle tumor Maignant Soft tissue profiferations, Muscle tissue, other to be specified) Soft tissue proliferations - Adipocytic Liposaroma, Undifferentiated Liposaroma, Undifferentiated Maignants Soft tissue profestions, Adipocytic, other (to be specified)	Primary utaneous perpheral T-cell Iymphoma, NOS Lymphocytic proliferations - Ge/Uk, other to be specified) Lymphocytic proliferations - B-Cell Primary utaneous marginal zone lymphoma Primary utaneous loide center hymphoma BRU+ mucoutaneous loider center hymphoma BRU+ mucoutaneous loider center hymphoma Intravacular large B-cell lymphoma
Melanocytic proliferations Atypical intraneptihelial melanocytic proliferation (AMP) Atypical Spitz tumor, Compound Atypical Spitz tumor, Compound Atypical Spitz tumor, Dermal (intradermal) Atypical profiterative nodukes in congenital melanocytic (Moderate or severe atypical CR mitotic activity) Superficial atypical melanocytic proliferation of uncertain significance (SAMPKS) Melanocytic tumor of uncertain malignant potential (VEETUMP) Atypical melanocytic neglesm todeterminate melanocytic norolferation, other (to be specified)	Epidermal proliferations Solar (activitic keratosis, NDS Solar (activitic keratosis, Hypertrophic Solar (activitic keratosis, Acartholytic Solar (activitic keratosis, Acartholytic Solar (activitic keratosis, Lichenoid Solar (activitic keratosis, Lichenoid Solar (activitic keratosis, other (to be specified) Solar (activitic keratosis, other (to be specified)		

FIGURE 7 Overview of the categories and terms under the 'malignant' (red) and 'indeterminate' (brown) super-categories.

inclusion of an entity in the 'indeterminate' category does not imply that these lesions have biological malignant potential, but rather difficulty in placing an entity into benign or malignant category.

The terms 'dysplastic nevus' and 'atypical nevus', whose definitions are varied,^{38–42} did not reach consensus by the experts following three Delphi rounds. In the first round 'dysplastic nevus', 'atypical nevus' and 'Clark nevus' were presented as three distinct terms under 'benign' 'melanocytic proliferations'; none surpassed the 75% threshold. Even in round III, the top two options—combine all three terms versus combine only 'atypical nevus' and 'dysplastic nevus'—received a tied vote. Finally, the steering committee, using a simple-majority vote, chose the more inclusive option of combining all three diagnoses to one ISIC-DX term 'Nevus, Atypical, NOS/Nevus, Dysplastic/Nevus, Clark'. This was thought to be the best approach given the overlapping use of the terms in labeling subsets of nevi, while being cognizant that these terms are not strict synonyms.

The proposed lexicon has limitations. First, we chose to restrict the scope of diagnostic terms for practicability. Future efforts may need to specifically enrich image data sets for rare diagnoses, and expand the taxonomy to include inflammatory or infectious disorders. Second, we chose to limit granularity for each diagnosis. While ancillary immunohistochemical and molecular markers may gain importance in distinguishing variants of diagnoses, Delphi members focused on the current clinical relevance of diagnostic entities. Eventually, we anticipate that a more molecularly informed ontology of diagnoses will lead to improved diagnostics labels. Lastly, the lexicon has not been tested prospectively. To understand the completeness, applicability and reproducibility of the taxonomy, we plan to study the prospective labeling of images submitted to the ISIC archive. We anticipate that the terms may need to be periodically updated.

In conclusion, we propose a publicly available, expert consensus-based taxonomy for diagnostic labeling of images of skin neoplasms. The ISIC-DX lexicon may benefit clinical communication, teaching and research and may improve AI training and interpretability. Further study and feedback from the dermatology community will be needed to evaluate the utility and the completeness of the lexicon.

AFFILIATIONS

¹The Kittner Skin Cancer Screening & Research Institute, Sheba Medical Center and Faculty of Medicine, Tel Aviv University, Tel Aviv, Israel

²Dermatology Service, Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, New York, USA

³Department of Dermatology-Venereology, University of Athens Medical School, Athens, Greece

⁴Department of Translational Research, Institut Curie, and UFR de Médecine, Université de Paris, Paris, France

⁵Department of Dermatology, University Hospital Zurich, Zurich, Switzerland ⁶Department of Dermatology, University of Arizona College of Medicine, and the University of Arizona Cancer Center Skin Cancer Institute, Tucson, Arizona, USA ⁷Department of Pathology and Laboratory Medicine, Hospital of the University of Pennsylvania, Philadelphia, Pennsylvania, USA

⁸Anatomic Pathology and Cytopathology Unit, Istituto Nazionale Tumori IRCCS Fondazione 'G. Pascale', Naples, Italy

⁹Department of Dermatology, UConn Health, Farmington, Connecticut, USA

¹⁰Department of Dermatology, University of Florida, Gainesville, Florida, USA
¹¹Departments of Dermatology and Pathology, Hospital Federal de Bonsucesso, Rio de Janeiro, Brazil

¹²First Department of Dermatology, Aristotle University, Thessaloniki, Greece
 ¹³Department of Dermatology, Brigham and Women's Hospital and Melanoma
 Program, Dana-Farber Cancer Institute, Boston, Massachusetts, USA
 ¹⁴Melanoma and Skin Cancer Unit and Department of Dermatology, Escuela de
 Medicina, Pontificia Universidad Católica de Chile, Santiago, Chile

¹⁵Department of Dermatology, University of Modena and Reggio Emilia, Modena and Dermatology Clinic, University of Rome, Rome, Italy

¹⁶The University of Queensland Diamantina Institute, University of Queensland, Dermatology Research Centre, Brisbane, Queensland, Australia

¹⁷ Ist Department of Dermatology-Venereology, Andreas Sygros Hospital, National and Kapodistrian University of Athens School of Medicine, Athens, Greece ¹⁸Dermatology Department, Hôpital Universitaire Lyon Sud, Hospices Civils de Lyon, Pierre-Bénite, France

¹⁹Department of Dermatology, Medical University of Vienna, Vienna, Austria

ACKNOWLEDGEMENTS

None.

FUNDING INFORMATION

This research was funded in part by a grant from the National Cancer Institute/National Institutes of Health (P30-CA008748) made to Memorial Sloan Kettering Cancer Center. Alon Scope was funded by the Israel Science Foundation (ISF-1546-16). H Peter Soyer holds an NHMRC MRFF Next Generation Clinical Researchers Program Practitioner Fellowship (APP1137127).

CONFLICT OF INTEREST STATEMENT

A Scope—Consulting fees (stated as not relevant for this publication) from Roche Pharmaceuticals (Israel) LTD; Payment or honoraria for lectures, presentations, speakers bureaus, article writing or educational events (stated as not relevant for this publication) from Janssen Israel. RP Braun-Advisory board (stated as not relevant for this publication) from Allmiral. MA Marchetti-Grants from Harry J. Lloyd Charitable Trust-Funding to institution for study unrelated to this work; Payment or honoraria for lectures, presentations, speakers bureaus, article writing or educational events from American Dermoscopy Meeting and Medscape. AA Marghoob-Royalties or licences from Up To Date and Informa publishing; Payment or honoraria for lectures, presentations, speakers bureaus, article writing or educational events from DermLite and FotoFinder; Receipt of equipment, materials, drugs, medical writing, gifts or other services from Heine, DermLite and FotoFinder. HP Soyer-Consulting fees (to institution) from Canfield Scientific Inc, MoleMap Australia Pty Ltd and Blaze Biosciences Inc; Patents planned, issued or pending for Microbiopsy Device; Stock or stock options-Shareholder MoleMap NZ Limited and Shareholder edermconsult GmbH. A Stratigos-Payment or honoraria for lectures, presentations, speakers bureaus, manuscript writing or educational events, made to institution, from BMS, Janssen Cilag, Sanofi, Genesis Pharma and Regeneron. L Thomas-Receipt of equipment, materials, drugs, medical writing, gifts or other services, made to institution, from Casio, 3 GEN, Heine, Fotofinder and C-Cube. H Kittler—Royalties or licences from Casio, Heine.

Meta-Optima, Barco; Payment or honoraria for lectures, presentations, speakers bureaus, article writing or educational events from Fotofinder, Heine, Almirall, Novartis, Pelpharam and Eli Lilly; Participation on a Data Safety Monitoring Board or Advisory Board for La Roche Posay. V Rotemberg—Grants—KL2 award to Dr. Rotemberg from NIH NCATS. AC Halpern—Consulting fees from Canfield Scientific Inc. and SciBase; stock or stock options from Mutual Funds. All the other authors, including K Liopyris, J Weber, RL Barnhill, CN Curiel-Lewandrowski, DE Elder, G Ferrara, JM Grant-Kels, T Jeunon, A Lallas, JY Lin, C Navarrete-Dechent and G Pellacani have no conflict of interest to disclose.

DATA AVAILABILITY STATEMENT

The data that supports the findings of this study are available at https://github.com/neuroelf/dermodelphi.

ETHICAL APPROVAL

Consistent with previous studies, an institutional review board approval was not applicable to this study, as it included no research subjects; all Delphi members were co-authors.

ORCID

Alon Scope https://orcid.org/0000-0001-9160-7411 Konstantinos Liopyris https://orcid. org/0000-0001-9566-8238 Aimilios Lallas https://orcid.org/0000-0002-7193-0964 Michael A. Marchetti https://orcid. org/0000-0002-1793-1851 Cristian Navarrete-Dechent https://orcid. org/0000-0003-4040-3640 Giovanni Pellacani https://orcid. org/0000-0002-7222-2951 Luc Thomas https://orcid.org/0000-0003-1995-2434

Allan C. Halpern [®] https://orcid.org/0000-0001-7320-1901

REFERENCES

- 1. Piepkorn MW, Barnhill RL, Elder DE, Knezevich SR, Carney PA, Reisch LM, et al. The MPATH-Dx reporting schema for melanocytic proliferations and melanoma. J Am Acad Dermatol. 2014;70:131–41.
- Esteva A, Kuprel B, Novoa RA, Ko J, Swetter SM, Blau HM, et al. Dermatologist-level classification of skin cancer with deep neural networks. Nature. 2017;542:115–8.
- 3. Haenssle HA, Fink C, Schneiderbauer R, Toberer F, Buhl T, Blum A, et al. Man against machine: diagnostic performance of a deep learning convolutional neural network for dermoscopic melanoma recognition in comparison to 58 dermatologists. Ann Oncol. 2018;29:1836–42.
- Tschandl P, Codella N, Akay BN, Argenziano G, Braun RP, Cabo H, et al. Comparison of the accuracy of human readers versus machinelearning algorithms for pigmented skin lesion classification: an open, web-based, international, diagnostic study. Lancet Oncol. 2019;20:938–47.
- 5. Daneshjou R, Smith MP, Sun MD, Rotemberg V, Zou J. Lack of transparency and potential bias in artificial intelligence data sets and algorithms: a scoping review. JAMA Dermatol. 2021;157:1362–9.
- Adamson AS, Smith A. Machine learning and health care disparities in dermatology. JAMA Dermatol. 2018;154:1247–8.

- 7. Marghoob AA. Standards in dermatologic imaging. JAMA Dermatol. 2015;151:819–21.
- Cassidy B, Kendrick C, Brodzicki A, Jaworek-Korjakowska J, Yap MH. Analysis of the ISIC image datasets: usage, benchmarks and recommendations. Med Image Anal. 2022;75:102305.
- 9. The ISIC archive. [cited 2024 Jan 13]. Available from: https://www. isic-archive.com
- Marchetti MA, Codella NCF, Dusza SW, Gutman DA, Helba B, Kalloo A, et al. Results of the 2016 International Skin Imaging Collaboration International Symposium on Biomedical Imaging challenge: comparison of the accuracy of computer algorithms to dermatologists for the diagnosis of melanoma from dermoscopic images. J Am Acad Dermatol. 2018;78:270–7.
- Marchetti MA, Liopyris K, Dusza SW, Codella NCF, Gutman DA, Helba B, et al. Computer algorithms show potential for improving dermatologists' accuracy to diagnose cutaneous melanoma: results of the International Skin Imaging Collaboration 2017. J Am Acad Dermatol. 2020;82:622–7.
- 12. Wall D, Meah N, York K, Bhoyrul B, Bokhari L, Abraham LS, et al. A global eDelphi exercise to identify Core domains and domain items for the development of a Global Registry of Alopecia Areata Disease Severity and Treatment Safety (GRASS). JAMA Dermatol. 2021;157:1–11.
- Ackerman AB, Böer A. Histopathologic diagnosis of adnexal epithelial neoplasms. Atlas and text. New York, NY: Ardor Scribendi; 2008.
- Elston D, Ferringer T, Ko CJ, Peckham S, High WA, DiCaudo DJ. Dermatopathology. 3rd ed. New York, NY: Elsevier; 2018.
- Elder DE, Piepkorn MW, Barnhill RL, Longton GM, Nelson HD, Knezevich SR, et al. Pathologist characteristics associated with accuracy and reproducibility of melanocytic skin lesion interpretation. J Am Acad Dermatol. 2018;79:52–9.
- Willemze R, Cerroni L, Kempf W, Berti E, Facchetti F, Swerdlow SH, et al. The 2018 update of the WHO-EORTC classification for primary cutaneous lymphomas. Blood. 2019;133:1703–14.
- 17. The Web's Free. ICD-10-CM/PCS medical coding reference. 2022. [cited 2024 Jan 13]. Available from: https://www.icd10data.com/
- Lott JP, Elmore JG, Zhao GA, Knezevich SR, Frederick PD, Reisch LM, et al. Evaluation of the Melanocytic Pathology Assessment Tool and Hierarchy for Diagnosis (MPATH-Dx) classification scheme for diagnosis of cutaneous melanocytic neoplasms: results from the International Melanoma Pathology Study Group. J Am Acad Dermatol. 2016;75:356–63.
- Lott JP, Boudreau DM, Barnhill RL, Weinstock MA, Knopp E, Piepkorn MW, et al. Population-based analysis of histologically confirmed melanocytic proliferations using natural language processing. JAMA Dermatol. 2018;154:24–9.
- Elmore JG, Barnhill RL, Elder DE, Longton GM, Pepe MS, Reisch LM, et al. Pathologists' diagnosis of invasive melanoma and melanocytic proliferations: observer accuracy and reproducibility study. BMJ. 2017;357:j2813.
- Boulkedid R, Abdoul H, Loustau M, Sibony O, Alberti C. Using and reporting the Delphi method for selecting healthcare quality indicators: a systematic review. PLoS One. 2011;6:e20476.
- Katragadda C, Finnane A, Soyer HP, Marghoob AA, Halpern A, Malvehy J, et al. Technique standards for skin lesion imaging: a Delphi consensus Statement. JAMA Dermatol. 2017;153:207–13.
- 23. IDDx Tutorial. [cited 2024 Jan 13]. Available from: https://www.youtu be.com/watch?v=E-TrVenvFjY
- Navarrete-Dechent C, Liopyris K, Molenda MA, Braun R, Curiel-Lewandrowski C, Dusza SW, et al. Human surface anatomy terminology for dermatology: a Delphi consensus from the International Skin Imaging Collaboration. J Eur Acad Dermatol Venereol. 2020;34:2659–63.
- 25. Repository of Delphi surveys and participant responses. [cited 2023 Nov 1]. Available from: https://github.com/neuroelf/dermodelphi
- ICD-10-CM codes. [cited 2024 Jan 13] Available from: https://www. icd10data.com/ICD10CM/Codes

- 27. SNOMED International codes. [cited 2024 Jan 13]. Available from: https://www.snomed.org/
 - Anand N, Edwards L, Baker LX, Chren MM, Wheless L. Validity of using billing codes from electronic health records to estimate skin cancer counts. JAMA Dermatol. 2021;157:1089–94.
 - Gupta S, Reintjes R, Trialonis-Suthakharan N. Analysis of the methodology of skin cancer incidence registration in German cancer registries. [cited 2024 Jan 13]. Available from: https://ace.amegroups. com/article/view/5156
 - Schulz S, Daumke P, Romacker M, López-García P. Representing oncology in datasets: standard or custom biomedical terminology? Inform Med Unlocked. 2019;15:100186.
 - Maley A, Patrawala S, Stoff B. Compliance with the College of American Pathologists Protocol for melanoma in synoptic and non-synoptic reports: a cross-sectional study. J Am Acad Dermatol. 2016;74:179–81.
 - 32. Radick AC, Reisch LM, Shucard HL, Piepkorn MW, Kerr KF, Elder DE, et al. Terminology for melanocytic skin lesions and the MPATH-Dx classification schema: a survey of dermatopathologists. J Cutan Pathol. 2021;48:733–8.
 - Katz I, O'Brien B, Clark S, Thompson CT, Schapiro B, Azzi A, et al. Assessment of a diagnostic classification system for management of lesions to exclude melanoma. JAMA Netw Open. 2021;4:e2134614.
 - Carney PA, Reisch LM, Piepkorn MW, Barnhill RL, Elder DE, Knezevich S, et al. Achieving consensus for the histopathologic diagnosis of melanocytic lesions: use of the modified Delphi method. J Cutan Pathol. 2016;43:830–7.
 - 35. Ackerman AB. Solar keratosis is squamous cell carcinoma. Arch Dermatol. 2003;139:1216-7.
 - 36. Fu W, Cockerell CJ. The actinic (solar) keratosis: a 21st-century perspective. Arch Dermatol. 2003;139:66–70.
 - 37. Marks R. Who benefits from calling a solar keratosis a squamous cell carcinoma? Br J Dermatol. 2006;155:23-6.

- 38. Elder D. The dysplastic nevus. Pathology. 1985;17:291-7.
- Ackerman AB. What naevus is dysplastic, a syndrome and the commonest precursor of malignant melanoma? A riddle and an answer. Histopathology. 1988;13:241–56.
- 40. Tucker MA, Halpern A, Holly EA, Hartge P, Elder DE, Sagebiel RW, et al. Clinically recognized dysplastic nevi: a central risk factor for cutaneous melanoma. JAMA. 1997;277:1439–44.
- Roesch A, Landthaler M, Vogt T. The dysplastic nevus. Separate entity, melanoma precursor or diagnostic dilemma? Hautarzt. 2003;54:871-83.
- 42. Hofmann-Wellenhof R, Marghoob AA, Zalaudek I. Large acquired nevus or dysplastic nevus: what's in the name of a nevus? JAMA Dermatol. 2016;152:623-4.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Scope A, Liopyris K, Weber J, Barnhill RL, Braun RP, Curiel-Lewandrowski CN, et al. International Skin Imaging Collaboration-Designated Diagnoses (ISIC-DX): Consensus terminology for lesion diagnostic labeling. J Eur Acad Dermatol Venereol. 2025;39:117–125. <u>https://doi.org/10.1111/jdv.20055</u>